

ADIPOENDOCRINOLOGY AND ADIPOPACRINOLOGY: EMERGING FIELDS OF STUDY ON THE ADIPOSE TISSUE

George N. Chaldakov¹, Ivan S. Stankulov¹, Marco Fiore², Mariyana G. Hristova³, Gorana Rančić⁴, Peter I. Ghenev⁵, and Pavel S. Pavlov¹

¹Division of Electron Microscopy, Department of Forensic Medicine, Medical University, Varna, Bulgaria, ²Institute of Neurobiology, CNR, Rome, Italy, ³Endocrinology Clinic, Medical University, Varna, Bulgaria, ⁴Institute of Histology and Embryology, Medical School, University of Niš, Niš, Yugoslavia, and ⁵Department of General and Clinical Pathology, Medical University, Varna, Bulgaria

Adipose tissue was conceived originally as merely passive, space-filling, fat storage tissue. However, in the last 10 years, investigations aimed at studying the endocrine secretion by adipose tissue have enjoyed explosive growth. The major secretory compartment of adipose tissue consists of adipocytes and stromal fibroblasts (adipofibroblasts). These cells secrete multiple bioactive molecules, conceptualized as adipokines or adipocytokines. Overall, this intellectual growth process framed an emerging field of study, adipoendocrinology. "Adipoendocrinology" connotes the study of the cellular and molecular biology of the endocrine function of adipose tissue in normal and diseased conditions. In humans, white adipose tissue is partitioned into a few large depots, including visceral and subcutaneous location, and many small depots, associated with heart, large blood vessels, major lymph nodes and other organs. The possibility that the endocrine secretory activity of large adipose depots may directly contribute to the elevated plasma levels of disease-associated adipokines has recently gained considerable attention. However, the paracrine secretory activity of organ-associated adipose tissue (the small adipose depots) has until now attracted little attention in the adipobiology of disease. Here we attempt to emphasize that studies aimed at evaluation of the paracrine secretion of organ-associated adipose tissue are becoming mandatory, since identification of the secreted molecules, particularly, adipokines, may yield clues to a possible transmission of pathogenic and/or protective stimuli, from the associated adipose tissue towards the interior of the associating organ. In this review we summarize most of the current information about adipoendocrinology and adipoparacrinology of various diseases.

Biomed Rev 2001; 12: 31-39.

INTRODUCTION

In recent years, the simple paradigm of adipocytes as merely fat storage cells is rapidly evolving into a complex paradigm of these cells as multipotential secretory cells. Arguably, the

most momentous changes that have occurred in the field of these studies have been the discovery of leptin, adipocyte-specific secretory protein, in the end of 1994 (1). Onwards, this seminal finding initiated a period of intense interest in the elucidation of the endocrine and paracrine roles of the

Received 1 November 2001 and accepted 5 December 2001.

Correspondence and reprint requests to Dr George N. Chaldakov, Division of Electron Microscopy, Department of Forensic Medicine, Medical University, BG-9002 Varna, Bulgaria. Tel.: 359 52 454 394; Fax: 359 52 222 584; E-mail: chaldakov@yahoo.com

adipose tissue and their potential involvement in the molecular mechanisms of obesity and related diseases. Overall, this intellectual grown process framed two emerging fields of study, adipoendocrinology and adipoparacrinology, respectively. These connote the study of the cellular and molecular biology of the endocrine/paracrine function of adipose tissue in normal and diseased conditions.

In humans, white adipose tissue (WAT) is partitioned into a few large depots, including visceral and subcutaneous location, and many small depots, associated with heart, large blood vessels, major lymph nodes (2-5), and even parasellar region in the brain (6), while, brown adipose tissue (BAT) is present around kidneys, adrenals, and aorta, and within the mediastinum and neck (7). Adipocytes are also located in bone marrow. Although adipose tissue has a similar uniform histological appearance in various parts of the human body, recent evidence suggests regional variations in the secretory protein production by adipose tissue (2-5,8).

Today we witness an increasing attention to the field of adipobiology, one of the most exciting examples being the rapidly growing interest in understanding the adipose tissue secretion and its involvement in the molecular mechanisms of a surprising variety of diseases besides obesity and related diseases. The major secretory compartment of adipose tissue consists of adipocytes and stromal fibroblasts (adipofibroblasts). Adipocytes are protein secreting cells that synthesize, (usually) store, and release proteins targeted to multiple secretory pathways (9-14). Adipofibroblasts are the major site of estrogen synthesis, mediated by the enzyme aromatase cytochrome P450 which converts androgens to estrogens (15-17). In effect, adipocytes and adipofibroblasts secrete a large number of multifunctional molecules, including cytokines, growth factors, enzymes, hormones, complement factors, and matrix proteins, conceptualized as adipokines (18-21) or adipocytokines (22-24). In addition to their importance in lipid, glucose and energy homeostasis, adipokines, using endocrine and/or paracrine pathways, exert important and often critical control over hematopoiesis (25-27), inflammation (25,27-30), hemostasis (31,32), complement activities (33), reproduction (34), angiogenesis (35,36), and feeding behavior (37,38). In a similar vein, adipokines now prove to be involved in mediating various diseased processes. Hence the development of adipoendocrinology and adipoparacrinology may indeed contribute to the understanding of pathogenesis and therapy of adipokine-associated diseases.

ADIPOSE TISSUE IS AN ENDOCRINE AND PARACRINE ORGAN PRODUCING ADIPOKINES

Since the isolation of the first known adipocyte-secreted protein, the serine protease adipsin (39), in 1987, the list of adipose tissue secretory proteins has greatly extended. Many, like

tumor necrosis factor-alpha (TNF- α) (31,40,41), IL-8 (42), macrophage colony-stimulating factor (43), leukemia inhibitory factor (44), and also leptin itself (28), appeared to be cytokines or chemokines (IL-8), whereas others appeared to be growth factors, enzymes, and hormones (15-24,31-33,45-64), identical to those secreted by various other tissues. In obesity, for example, the plasminogen activator inhibitor-1 (PAI-1) secretory capacity of adipocytes may even exceed that of PAI-1-producing cells in other tissues (31). Likewise, in postmenopausal women, the estrogen production in whole body adipose tissue may be enough to effectively increase circulating levels of estrogens (15,16). Conceptually, to embody the secretory production by adipose tissue, the terms "adipokines" and "adipocytokines" were designated. Accordingly, their counterparts derived from nonadipose tissue sources are not in the scope of the present article. Note that mast cells, being an essential component of the adipose tissue, may also contribute to its secretory potential. Information about mast cell secretion and mast cell involvement in metabolic disorders may be found elsewhere (65-67).

Among adipokines discovered until now, leptin (from Greek *leptos*, means thin), encoded by the obese (*ob*) gene (1,25,26,28,30,34-38), and recently adiponectin (synonyms: Acrp30, adipocyte complement-related protein of 30 kD; apM1 protein, encoded by adipose most abundant gene transcript 1; AdipoQ; GBP28, gelatin-binding protein of 28 kD) (22-24,68-71) have received intense scientific coverage. Other adipokines such as PAI-1, tissue factor (TF), TNF- α and transforming growth factor-beta (TGF- β) (31), and recently resistin (synonyms: ADSF, adipocyte secreted factor; FIZZ3, found in inflammatory zone 3) and resistin-like molecules (58,59) have also attracted great attention. A list of adipokines is presented in Table 1.

ADIPOBIOLOGY OF ADIPOKINE-ASSOCIATED DISEASES

As indicated in the Introduction, adipose tissue is partitioned into a few large depots, including subcutaneous and visceral location, and many small depots, associated with heart, large blood vessels, major lymph nodes, bone marrow, kidneys, adrenal glands, and even the brain (2-7). All these fat depots are potential sources of adipokines. The possibility that the endocrine secretory activity of large adipose depots may directly contribute to the elevated plasma levels of disease-associated adipokines has recently gained considerable attention (21-26,31,32,52-56,72-84). However, the paracrine secretory activity of the small adipose depots has until now attracted little attention in the adipobiology of disease. If signals can, *via* endocrine (systemic) pathway, be targeted from the large adipose depots through the bloodstream towards many organs in the body, and hence lead to various metabolic and cardiovascular disorders, then why not look for similar, but

Table 1. A selected list of adipokines

Adipokines
Adipsin
Leptin
Adiponectin (ADNC)
Plasminogen activator inhibitor-1 (PAI-1)
Tissue factor (TF)
Tumor necrosis factor- α (TNF- α)
Transforming growth factor- β (TGF- β)
Macrophage colony-stimulating factor (MCSF)
Nerve growth factor (NGF)
Estrogens
Interleukin-6 (IL-6)
Interleukin-8 (IL-8)
Leukemia inhibitory factor
Lipoprotein lipase (LPL)
Acylation stimulating protein
Angiotensin II
Cathepsin D/G
Metallothionein-1 (MT-1)
Bone morphogenetic protein GDF-2
Found in inflammatory zone 1 (FIZZ1)
Resistin (FIZZ3)
Heparin-binding epidermal growth factor (HB-EGF)
Osteonectin (SPARC)*
Hevin

* SPARC, secretory protein, acidic and rich in cysteine.

paracrine (local) reactions from the organ-associated adipose tissue represented by the small adipose depots? It was emphasized that studies aimed at evaluation of the molecular composition of the organ-associated adipose tissue become mandatory, since identification of these molecules, particularly, adipokines, may yield clues to a possible transmission of pathogenic and/or protective stimuli, from the associated adipose tissue towards the interior of the associating organ (2-4,18,19,21,85-88). In a similar vein, this defines a new field of study, adipoparacrinology. An intriguing example is the subepicardial adipose tissue (SEAT) that is conjunctioned to the adventitia of the most atherosclerosis-prone portions of the coronary artery, that is, the most proximal part of its left anterior descending (LAD) branch. In 1933, Smith and Willius (cited in 4) have pointed out a functional relationship between the SEAT and the LAD coronary artery, and stated

that SEAT is “not a passive storehouse for fat”. From metabolic point of view, the principle difference between SEAT and adipose tissue elsewhere in the body is its greater capacity for free fatty acid release and uptake, thus acting as a local energy supply for epimyocardium and coronary arteries and/or as a buffer against toxic levels of free fatty acids (4). Neglected for nearly 70 years, the possible involvement of SEAT in atherogenesis has been, at long last, currently addressed (18,86-88). Specifically, recent findings demonstrate that (i) the portion of the LAD coronary artery running in the SEAT develops atherosclerotic lesions, while the portion running in the myocardium is free of atherosclerotic lesions (86), and (ii) the “atherosclerotic” SEAT expresses an increased amount of nerve growth factor (NGF) and mast cells in human coronary atherosclerosis (21,77,87,88). Other examples of a potential pathogenic involvement of organ-associated adipose tissue include breast cancer (15,16), thyroid-associated ophthalmopathy (89,90), Crohn’s disease (91), HIV-associated fat redistribution syndrome (see 2-5), and pheochromocytoma (92). It is also worth noting that a recent work shows that subcutaneous adipocytes express NGF, supposedly involved in the process of wound healing (93), whereas topically applied NGF exerts beneficial effects in human skin ulcers (94,95). Another adipoparacrinological challenge might be studying the potential role of parasellar and epidural adipose tissue in pituitary gland and spinal cord pathology, respectively. Together, these findings strongly suggest that to further elucidate the potential physiological and pathogenic importance of organ-associated adipose tissue, we should no longer, as hitherto, cut it from the respective organ, but keep it attached and in place, and subject to thorough examination. Applying such an adipoprotective action, recent studies reported important findings on (i) the participation of perinodal adipose tissue (major lymph node-associated adipose tissue) in immune responses (2-5), and (ii) the modulation of contractile responsiveness of aorta by perivascular adipose tissue (96; see also 97 about leptin-induced nitric oxide release).

Figure 1 schematically illustrates endocrine (systemic) and paracrine (local) approach in studying the adipokine-associated diseases, whereas Table 2 presents a selective list of these diseases.

CONCLUSION

At birth, the average-size infant has approximately five billion adipocytes, whereas - approximately 80 billion in adult (6). Adding to them billions of fibroblasts and mast cells, makes the whole body adipose tissue, particularly WAT, a major human’s secretory organ, topologically organized as visceral and subcutaneous adipose tissue (the large adipose depots) and organ-associated adipose tissue (the small adipose depots).

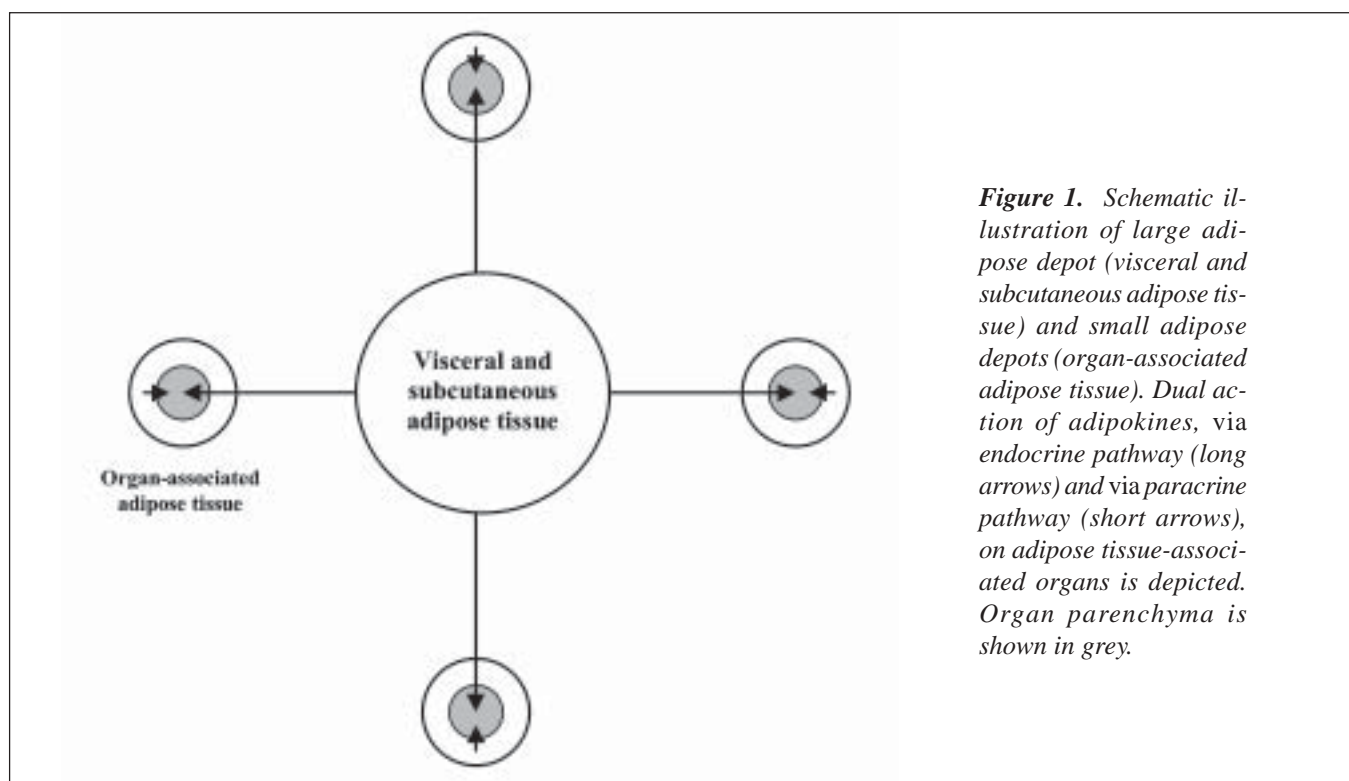


Table 2. Selected list of adipokine-associated diseases*

<u>Diseases</u>	<u>Adipokines/Endocrine secretion</u>
Obesity	Leptin, ADNC, TNF- α , IL-6, PAI-1, TF, TGF- β , Resistin
Type II diabetes	ADNC, Leptin, TNF- α , PAI-1, TF, TGF- β
Metabolic syndrome	ADNC, TNF- α , IL-6, PAI-1, Leptin, NGF
Cardiovascular disease	ADNC, PAI-1, TF, TNF- α , TGF- β , Leptin, SPARC, HB-EGF, MT-1 (?)
Hypertension	Leptin, Angiotensin II
Hypertensive retinopathy	Leptin
Myocardial infarction	Leptin, ADNC (?)
Hemorrhagic stroke	Leptin
<u>Diseases</u>	<u>Adipokines/Paracrine secretion</u>
Coronary atherosclerosis	NGF
Breast cancer	Estrogens, IL-1,-6, TNF- α , NGF, FIZZ1 (?)
Crohn's disease	TNF- α
Skin wound	NGF
Thyroid-associated ophthalmopathy	IL-1, TNF- α
Osteoporosis	LPL, Estrogens
Phaeochromocytoma	?

* For abbreviations see Table 1.

Further study of both visceral and subcutaneous adipose tissue should certainly be continued. However, we should pay more attention to further elucidate the role of organ-associated adipose tissue in health and disease.

In recent years, the discovery that adipose tissue cells secrete a plethora of disease-related adipokines led to the development of major new insights into adipobiology at both the basic and clinical levels. For instance, a cluster of adipokines consisted of PAI-1, TF, TGF- β , TNF- α , IL-6, adiponectin, leptin, resistin, HB-EGF, NGF, osteonectin/SPARC, and MT-1 is differentially linked to obesity and related diseases. Since the actions of adipokines, like those of their relatives derived from nonadipose tissue sources, are complex and diverse, we need to design many new studies to determine how these bioactive molecules can, under different conditions, both promote and suppress various processes triggered by obesity, insulin resistance, and inflammation. The present challenge is thus to cultivate much a new thinking about how can we make adipokines work for the therapeutic benefit of patients. Although the development of novel therapies such as gene therapy, recombinant protein therapy, and stem cell-based therapy attracts great attention now, the search for small molecules as drugs remains prospective strategies in adipopharmacology. For instance, further studies may specifically upregulate some hidden, adipokine-associated actions of drugs that are already in clinical practice, like pentoxifylline, peroxisome proliferator-activated receptor-gamma agonists, and tamoxifen (41, 98-100). In this context, it is worth reminding that aspirin kept hidden its antiplatelet therapeutic action until the discovery of prostanoids, and so did, until the discovery of cytokines, pentoxifylline about its anti-TNF- α , antiinflammatory potential, and recently, antidepressants' action was related to the elevations of neurotrophin levels in the brain (101). Hence we could search some "old" drugs to disclose their adipokine-mediated therapeutic potential.

Finally, understanding the molecular biology and pharmacology of adipose tissue secretion could lead to novel ways to prevent and treat adipokine-associated diseases. Further progress in adipoendocrinology and adipoparacrinology holds much promise for the achievement of that goal.

ACKNOWLEDGMENTS

We thank Dr Luigi Aloe, Institute of Neurobiology, National Research Council, Rome, Italy for the support and the collaboration. Creative reading of the manuscript by Dr Anton B. Tonchev at Kanazawa University Medical School, Kanazawa, Japan is highly appreciated.

REFERENCES

- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994; 372: 425-432.
- Pond CM. Interactions between adipose tissue and the immune system. *Proc Nutr Soc* 1996; 55: 111-126.
- Pond CM. *The Fats of Life*. Cambridge University Press, Cambridge. 1998.
- Pond CM. Physiological specialisation of adipose tissue. *Prog Lipid Res* 1999; 38: 225-248.
- Pond CM. Adipose tissue, the anatomists' Cinderella, goes to the ball at last, and meets some influential partners. *Postgrad Med J* 2000; 76: 671-673.
- Weninger WJ, Pramhas D. Compartments of the adult parasellar region. *J Anat* 2000; 197 (Pt 4): 681-686.
- Brooks JJ, Perosio PM. Adipose tissue. In: Stephen R, Strenberg S, editors. *Histology for Pathologists*. Raven Press, New York. 1992; 33-60.
- Arner P. Regional differences in protein production by human adipose tissue. *Biochem Soc Trans* 2001; 29 (Pt 2): 72-75.
- Park JW, Blanchette-Mackie EJ, Scow RO, Brefeldin A enables synthesis of active lipoprotein lipase in cld/cld and castanospermine-treated mouse brown adipocytes via translocation of Golgi components to endoplasmic reticulum. *Biochem J* 1996; 317 (Pt 1): 125-134.
- Yang CZ, Mueckler M. ADP-ribosylation factor 6 (ARF6) defines two insulin-regulated secretory pathways in adipocytes. *J Biol Chem* 1999; 274: 25297-25300.
- Watson RT, Pessin JE. Functional cooperation of two independent targeting domains in syntaxin 6 is required for its efficient localization in the trans-Golgi network of 3T3-L1 adipocytes. *J Biol Chem* 2000; 275: 1261-1268.
- Millar CA, Meerloo T, Martin S, Hickson GR, Shimwell NJ, Wakelam MJ, *et al*. Adipsin and the glucose transporter GLUT4 traffic to the cell surface via independent pathways in adipocytes. *Traffic* 2000; 1: 141-151.
- Roh C, Roduit R, Thorens B, Fried S, Kandror KV. Lipoprotein lipase and leptin are accumulated in different secretory compartments in rat adipocytes. *J Biol Chem* 2001; 276: 35990-35994.
- Halleux CM, Takahashi M, Delporte ML, Detry R, Funahashi T, Matsuzawa Y, *et al*. Secretion of adiponectin and regulation of apM1 gene expression in human visceral adipose tissue. *Biochem Biophys Res Commun* 2001; 288: 1102-1107.
- Bulun SE, Sharda G, Rink J, Sharma S, Simpson ER. Distribution of aromatase P450 transcripts and adipose fibroblasts in the human breast *J Clin Endocrinol Metab* 1996; 81: 1273-1277.
- Agarwal VR, Bulun SE, Leftch M, Rohrich R, Simpson ER. Use of alternative promoters to express the aromatase cytochrome P450 (CYP19) gene in breast adipose tissues of cancer-free and breast cancer patients. *J Clin Endocrinol Metab* 1996; 81: 3843-3849.

17. Cannady WE, Brann DW, Mahesh VB. The potential role of periovarian fat and leptin in initiation of puberty in the immature rat. *Int J Obes Relat Metab Disord* 2000; 24 (Suppl 2): S146-S147.
18. Chaldakov GN, Fiore M, Ghenev PI, Stankulov IS, Aloe L. Atherosclerotic lesions: possible interactive involvement of intima, adventitia and associated adipose tissue. *Int Med J* 2000; 7: 43-49.
19. Chaldakov GN, Fiore M, Hristova M, Aloe L. Cell biology and pharmacology of adipose tissue secretion. *FABAD J Pharm Sci* 2000; 25: 181-191.
20. Chaldakov G, Fiore M, Hristova M, Stankulov I, Triaca V, Ghenev P, *et al.* Adipose tissue-secreted molecules (adipokines): neuroimmune implications. *Clin Appl Immunol Invest* 2000; 1: 11-16.
21. Chaldakov GN, Fiore M, Stankulov IS, Hristova M, Antonelli A, Manni L, *et al.* Neurotrophins, adipokines, and mast cells in human cardiovascular and metabolic pathology: A new road ahead. *Scr Sci Med* 2001; 33: 101-104.
22. Matsuzawa Y, Funahashi T, Nakamura T. Molecular mechanism of metabolic syndrome X: contribution of adipocytokines, adipocyte-derived bioactive substances. *Ann NY Acad Sci* 1999; 892: 146-154.
23. Funahashi T, Nakamura T, Shimomura I, Maeda K, Kuriyama H, Takahashi M, *et al.* Role of adipocytokines on the pathogenesis of atherosclerosis in visceral obesity. *Intern Med* 1999; 38: 202-206.
24. Ouchi N, Kihara S, Arita Y, Okamoto Y, Maeda K, Kuriyama H, *et al.* Adiponectin, an adipocyte-derived plasma protein, inhibits endothelial NF- κ B signaling through a cAMP-dependent pathway. *Circulation* 2000; 102: 1296-1301.
25. Fantuzzi G, Faggioni R. Leptin in the regulation of immunity, inflammation, and hematopoiesis. *J Leuk Biol* 2000; 68: 437-446.
26. Ozata M, Ozdemir IC, Licinio J. Human leptin deficiency caused by a missense mutation: multiple endocrine defects, decreased sympathetic tone, and immune system dysfunction indicate new targets for leptin action, greater central than peripheral resistance to the effects of leptin, and spontaneous correction of leptin-mediated defects. *J Clin Endocrinol Metab* 1999; 84: 3686-3695.
27. Yokata T, Oritani K, Takahashi T, Ishikawa J, Matsuyama A, Ouchi N, *et al.* Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the function of macrophages. *Blood* 2000; 96: 1723-1732.
28. Lofifreda S, Yang Q, Lin HZ, Karp CL, Brengman ML, Wang DJ, *et al.* Leptin regulates proinflammatory immune responses. *FASEB J* 1998; 12: 57-65.
29. Ouchi N, Kihara S, Arita Y, Maeda K, Kuriyama H, Okamoto Y, *et al.* Novel modulator for endothelial adhesion molecules: adipocyte-derived plasma protein adiponectin. *Circulation* 1999; 100: 2473-2476.
30. Zarkesh-Esfahani H, Pockley G, Metcalfe RA, Bidlingmaier M, Wu Z, Ajami A, *et al.* High-dose leptin activates human leukocytes via receptor expression on monocytes. *J Immunol* 2001; 167: 4593-4599.
31. Loskutoff DJ, Fujisawa K, Samad F. The fat mouse. A powerful genetic model to study hemostatic gene expression in obesity/NIDDM. *Ann NY Acad Sci* 2000; 902: 272-282.
32. Mutch NJ, Wilson HM, Booth NA. Plasminogen activator inhibitor-1 and haemostasis in obesity. *Proc Nutr Soc* 2001; 60: 341-347.
33. Choy LN, Spiegelman BM. Regulation of alternative pathway activation and C3a production by adipose cells. *Obes Res* 1996; 4: 521-532.
34. Hileman SM, Pierroz DD, Flier JS. Leptin, nutrition, and reproduction: timing is everything. *J Clin Endocrinol Metab* 2000; 85: 804-807.
35. Kang SM, Kwon HM, Hong BK, Kim D, Kim IJ, Choi EY, *et al.* Expression of leptin receptor (Ob-R) in human atherosclerotic lesions: potential role in intimal neovascularization. *Yonsei Med J* 2000; 41: 68-75.
36. Park HY, Kwon HM, Lira HJ, Hong BK, Lee JY, Park BE, *et al.* Potential role of leptin in angiogenesis: leptin induces endothelial cell proliferation and expression of matrix metalloproteinases in vivo and in vitro. *Exp Mol Med* 2001; 33: 95-102.
37. Friedman JM, Halaas JL. Leptin and the regulation of body weight in mammals. *Nature* 1998; 395: 763-770.
38. Flier JS. Clinical review 94: What's in a name? In search of leptin's physiologic role. *J Clin Endocrinol Metab* 1998; 83: 1407-1413.
39. Cook KS, Min HY, Johnson D, Chaplinsky RJ, Flier JS, Hunt CR, *et al.* Adipsin: a circulating serine protease homologue secreted by adipose tissue and sciatic nerve. *Science* 1987; 237: 402-405.
40. Zhang HH, Kumar S, Barnett AH, Eggo MC. Tumour necrosis factor- α exerts dual effects on human adipose leptin synthesis and release. *Mol Cell Endocrinol* 2000; 159: 79-88.
41. Gigolini M, Tonoli M, Borgato L, Frigotto L, Manzato F, Zemman S, *et al.* Expression of plasminogen activator inhibitor-1 in human adipose tissue: a role of tumor necrosis factor- α ? *Atherosclerosis* 1999; 143: 81-90.
42. Bruun JM, Pedersen SB, Richelsen B. Regulation of interleukin 8 production and gene expression in human adipose tissue in vitro. *J Clin Endocrinol Metab* 2001; 86: 1267-1273.
43. Levine JA, Jensen MD, Eberhardt NL, O'Brien T. Adipocyte macrophage colony-stimulating factor is a

- mediator of adipose tissue growth. *J Clin Invest* 1998; 101: 1557-1564.
44. Aubert J, Dessolin S, Belmonte N, Li M, McKenzie FR, Staccini L, *et al.* Leukemia inhibitory factor and its receptor promote adipocyte differentiation via the mitogen-activated protein kinase cascade. *J Biol Chem* 1999; 274: 24965-24972.
 45. Lundgren CH, Brown SL, Nordt TK, Sobel BE, Fujii S. Elaboration of type-1 plasminogen activator inhibitor from adipocytes. A potential pathogenetic link between obesity and cardiovascular disease. *Circulation* 1996; 93: 106-110.
 46. Maeda K, Okubo K, Shimomura I, Mizumo K, Matsuzawa Y, Matsubara K. Analysis of an expression profile of genes in the human adipose tissue. *Gene* 1997; 190: 227-235.
 47. Samad F, Pandey M, Loskutoff DJ. Tissue factor gene expression in the adipose tissues of obese mice. *Proc Natl Acad Sci USA* 1998; 95: 7591-7596.
 48. Morange PE, Alessi MC, Verdier M, Casanova D, Magalon G, Juhan-Vague I. PAI-1 produced ex vivo by human adipose tissue is relevant to PAI-1 blood level. *Arterioscler Thromb Vasc Biol* 1999; 19: 1361-1365.
 49. Samad F, Uysal KT, Wiesbrock SM, Pandey M, Hotamisligil GS, Loskutoff DJ. Tumor necrosis factor alpha is a key component in the obesity-linked elevation of plasminogen activator inhibitor 1. *Proc Natl Acad Sci USA* 1999; 96: 6902-6907.
 50. Claeskens A, Ongenaes N, Neefe JM, Cheyns P, Kaijen P, Cools M, *et al.* Hevin is down-regulated in many cancers and is a negative regulator of cell growth and proliferation. *Br J Cancer* 2000; 82: 1123-1130.
 51. Holcomb IN, Kabakoff RC, Chan B, Baker TW, Gumey A, Henzel W, *et al.* FIZZ1, a novel cyteine-rich secreted protein associated with pulmonary inflammation, defines a new gene family. *EMBO J.* 2000; 19: 4046-4055.
 52. Trayhurn P, Duncan JS, Wood AM, Beattie JH. Regulation of metallothionein gene expression and secretion in rat adipocytes differentiated from preadipocytes in primary culture. *Horm Metab Res* 2000; 32: 542-547.
 53. Göbel H, van der Wal A, Teeling P, van der Loos C, Becker AE. Metallothionein in human atherosclerotic lesions: a scavenger mechanism for reactive oxygen species in the plaque? *Virchows Arch* 2000; 437: 528-533.
 54. Trayhurn P, Beattie JH. Physiological role of adipose tissue: white adipose tissue as an endocrine and secretory organ. *Proc Nutr Soc* 2001; 60: 329-339.
 55. Witthuhn BA, Bernlohr DA. Upregulation of bone morphogenetic protein GDF-2/Vgr-2 expression in adipose tissue of FABP4/aP2 null mice. *Cytokines* 2001; 14: 129-135.
 56. Tartare-Deckert S, Chavey C, Monthouel MN, Gautier N, Van Obberghen E. The matricellular protein SPARC/osteonectin as a newly identified factor up-regulated in obesity. *J Biol Chem* 2001; 276: 22231-22237.
 57. Hausman DB, DiGirolamo M, Bartness TJ, Hausman GJ, Martin RJ. The biology of white adipocyte proliferation. *Obes Rev* 2001; 2: 239-254.
 58. Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, *et al.* The hormone resistin links obesity to diabetes. *Nature* 2001; 409: 307-312.
 59. Steppan CM, Brown EJ, Wright CM, Bhat S, Banerjee RR, Dai CY *et al.* A family of tissue-specific resistin-like molecules. *Proc Natl Acad Sci USA* 2001; 98: 502-506.
 60. Karlsson C, Lindell K, Ottosson M, Sjöström L, Carlson B, Carlsson LMS. Human adipose tissue expresses angiotensinogen and enzyme required for its conversion to angiotensin II. *J Clin Endocrinol Metab* 1998; 83: 3925-3929.
 61. Engeli S, Gorzelniak K, Kreutz R, Runkel N, Distler A, Sharma AM. Co-expression of renin-angiotensin system genes in human adipose tissue. *J Hypertens* 1999; 17: 555-560.
 62. Sasano H, Ozaki M. Aromatase expression and its localization in human breast cancer. *J Steroid Biochem Mol Biol* 1997; 61: 293-298.
 63. Miller WR. Biology of aromatase inhibitors: pharmacology and endocrinology within the breast. *Endocr Relat Cancer* 1999; 6: 187-195.
 64. Bhatnagar AS, Brodie AM, Long BJ, Evans DB, Miller WR. Intracellular aromatase and its relevance to the pharmacological efficacy of aromatase inhibitors. *J Steroid Biochem Mol Biol* 2001; 76: 199-202.
 65. Ruger BM, Hasan Q, Greenhill NS, Davis PF, Dunbar PR, Neale TJ. Mast cells and type VIII collagen in human diabetic nephropathy. *Diabetologia* 1996; 39: 1215-1222.
 66. Hatanaka K, Tanishita H, Ishibashi-Ueda H, Yamamoto A. Hyperlipidemia in mast cell-deficient W/WV mice. *Biochem Biophys Acta* 1986; 878: 440-445.
 67. Hristova M, Aloe L, Ghenev PI, Fiore M, Chaldakov GN. Leptin and mast cells: a novel adipimmune link. *Turk J Med Sci* 2001; 31: 581-583.
 68. Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to Clq, produced exclusively in adipocytes. *J Biol Chem* 1995; 270: 26746-26749.
 69. Maeda K, Okudo K, Shimomura I, Funahashi T, Matsuzawa Y, Matsubara K. cDNA cloning and expression of a novel adipose specific collage-like factor, apM1 (AdiPose Most abundant Gene transcript 1). *Biochem Biophys Res Commun* 1996; 221: 286-289.
 70. Hu E, Liang P, Spiegelman BM. AdipoQ is a novel adipose-specific gene dysregulated in obesity. *J Biol Chem* 1996; 271: 10697-10703.
 71. Nakano Y, Tobe T, Choi-Miura NH, Mazda T, Tomita M. Isolation and characterization of GBP28, a novel gelatin-

- binding protein purified from human plasma. *J Biochem (Tokyo)* 1996; 120: 803-812.
72. Schaffler A, Orso E, Palitzsch KD, Buchler C, Drobnik W, Furst A, *et al.* The human apM-1, an adipose-specific gene linked to the family of TNF's and to genes expressed in activated T cells, is mapped to chromosome 1q21.3-q23, a susceptibility locus identified for familial combined hyperlipidaemia (FCH). *Biochem Biophys Res Commun* 1999; 260: 416-425.
 73. Uckaya G, Ozata M, Sonmer A, Kinalp C, Eyileten T, Bingol N, *et al.* Plasma leptin levels strongly correlate with plasma renin activity in patients with essential hypertension. *Horm Metab Res* 1999; 31: 435-38.
 74. Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J-I, *et al.* Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 1999; 257: 79-83.
 75. Yanovski JA, Yanovski SZ. Recent advances in basic obesity research. *JAMA* 1999; 282: 1504-1506.
 76. Morange PE, Lijnen HR, Alessi MC, Kopp F, Collen D, Juhan-Vague P. Influence of PAI-1 on adipose tissue growth and metabolic parameters in a murine model of diet-induced obesity. *Arterioscler Thromb Vasc Biol* 2000; 20:1150-1154.
 77. Chaldakov GN, Fiore M, Stankulov IS, Hristova M, Antonelli A, Manni L, *et al.* NGF, BDNF, leptin, and mast cells in human coronary atherosclerosis and metabolic syndrome. *Arch Physiol Biochem* 2001; 109: 357-360.
 78. Uckaya G, Ozata M, Sonmez A, Kinalp C, Eyileten T, Bingol N, *et al.* Is leptin associated with hypertensive retinopathy. *J Clin Endocrinol Metab* 2000; 85: 683-687.
 79. Statnick MA, Beavers LS, Conner LJ, Corominola H, Johnson D, Hammond CD, *et al.* Decreased expression of apMI in omental and subcutaneous adipose tissue of humans with type 2 diabetes. *Int J Exp Diabetes Res* 2000; 1: 81-88.
 80. Hotta K, Funahashi T, Arita Y, Takahashi M, Matsuda M, Okamoto Y, *et al.* Plasma concentration of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol* 2000; 20: 1595-1599.
 81. Schubring C, Blum WF, Kratzsch J, Deutscher J, Kiess W. Leptin, the ob gene product, in female health and disease. *Eur Obstet Gynecol Reprod Biol* 2000; 88: 121-127.
 82. Comuzzie AG, Funahashi T, Sonnenberg G, Martin LJ, Jacob HJ, Black AE, *et al.* The genetic basis of plasma variation in adiponectin, a global endophenotype for obesity and the metabolic syndrome. *J Clin Endocrinol Metab* 2001; 86: 4321-4325.
 83. Weyer C, Funahashi T, Tanaka S, Hotta K, Matsuzawa Y, Pratley RE, *et al.* Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metab* 2001; 86: 1930-1935.
 84. Ouchi N, Kihara S, Arita Y, Nishida M, Matsuyama A, Okamoto Y, *et al.* Adipocyte-derived plasma protein, adiponectin, suppresses lipid accumulation and class A scavenger receptor expression in human monocyte-derived macrophages. *Circulation* 2001; 103: 1057-1063.
 85. Chaldakov GN, Fiore M, Ghenev PI, Stankulov IS, Angelucci F, Pavlov PS, *et al.* Conceptual novelties in atherogenesis: smooth muscle cells, adventitia, and adipose tissue. *Biomed Rev* 2000; 11: 63-67.
 86. Ishii T, Asuwa N, Masuda S, Ishikawa Y. The effects of a myocardial bridge on coronary atherosclerosis and ischemia. *J Pathol* 1998;185: 4-9.
 87. Chaldakov GN, Stankulov IS, Aloe L. Subepicardial adipose tissue in human coronary atherosclerosis: another neglected phenomenon. *Atherosclerosis* 2001; 154: 237-238.
 88. Chaldakov GN, Stankulov IS, Fiore M, Ghenev PI, Aloe L. Nerve growth factor levels and mast cell distribution in human coronary atherosclerosis. *Atherosclerosis* 2001; 159: 57-66.
 89. Hiromatsu Y, Yang D, Bednarczuk T, Miyake I, Nonaka K, Inoue Y. Cytokine profiles in eye muscle tissue and orbital fat tissue from patients with thyroid-associated ophthalmopathy. *J Clin Endocrinol Metab* 2000; 85: 1194-1199.
 90. Valyasevi RW, Jyonouchi SC, Dutton CM, Munsakul N, Bahn RS. Effect of tumor necrosis factor- α , interferon- γ , and transforming growth factor- β on adipogenesis and expression of thyrotropin receptor in human orbital preadipocyte fibroblasts. *J Clin Endocrinol Metab* 2001; 86: 903-908.
 91. Desreumaux P, Ernts O, Geboes K, Gambiez L, Berrebi D, Muller-Alouf H, *et al.* Inflammatory alterations in mesenteric adipose tissue in Crohn's disease. *Gastroenterology* 1999; 117: 73-81.
 92. Lean ME, James WP, Jennings G, Trayhurn P. Brown adipose tissue in patients with pheochromocytoma. *Int J Obes* 1986; 10: 219-227.
 93. Hasan W., Zhang R, Liu M, Warn JD, Smith PG. Coordinated expression of NGF and alpha-smooth muscle actin mRNA and protein in cutaneous wound tissue of developing and adult rats. *Cell Tissue Res* 2000; 300: 97-109.
 94. Levi-Montalcini R. *The Saga of the Nerve Growth Factor. Preliminary Studies, Discovery, Further Development.* World Scientific, Singapore, 1997.
 95. Aloe L, Tirassa P, Bracci-Laudiero L. Nerve growth factor in neurological and non-neurological diseases: basic findings and emerging pharmaceutical perspectives. *Curr Pharm Des* 2001; 7: 113-123.

96. Soltis EE, Cassis LA. Influence of perivascular adipose tissue on rat aortic smooth muscle responsiveness. *Clin Exp Hypertens A* 1991; 13: 277-296.
97. McCann SM, Yu WH, Karanth S, Mastronardi CA. Control of the release of leptin, tumor necrosis factor-alpha and nitric oxide by the central nervous system. [abstract]. *Biomed Rev* 2001; 12: 66.
98. Maeda N, Takahashi M, Funahashi T, Kihara S, Nishizawa H, Kishida K, *et al.* PPAR gamma ligands increase expression and plasma concentration of adiponectin, an adipose-derived protein. *Diabetes* 2001; 50: 2094-2099.
99. Rubin GL, Zhao Y, Kalus AM, Simpson ER. Peroxisome proliferator-activated receptor gamma ligands inhibit estrogen biosynthesis in human breast adipose tissue: possible implications for breast cancer therapy. *Cancer Res* 2000; 60: 1604-1608.
100. Chiarenza A, Lazarovici P, Lempereur L, Cantarella G, Bianchi A, Bernardini R. Tamoxifen inhibits nerve growth factor-induced proliferation of human breast cancerous cell line MCF-7. *Cancer Res* 2001; 61: 3002-3008.
101. Altar CA. Neurotrophins and depression. *Trends Pharmacol Sci* 1999; 20: 59-61.