

NEURONAL RECOVERY AFTER PERIPHERAL TRAUMATIC LESIONS OF THE FACIAL MOTOR NERVE

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SUMMARY

• *The present work summarizes recent data on the responses in the facial nucleus to experimental lesion on the extratemporal part of the facial nerve. It includes a description of all so far reported reactions to axotomy in motoneurons, microglia and astroglia at cellular and molecular level, together with an attempt for a comprehensible interpretation of their importance. The brief review on the experimental and clinical results of the reinnervation is followed by some original results from our experience to improve neuronal regeneration with peroral administration of the calcium antagonist nimodipin.*

INTRODUCTION

• The facial nerve is the most emotive nerve in the human body. Its motor (mimic) function accounts for one's facial tone, voluntary facial movement, and involuntary emotional expression of the face, i.e. the aesthetic appearance of an individual, which is so important for social life. That is why the dysfunction of the facial nerve in combination with the grotesque disfigurement may often lead to a psychological incapacitation (1-3).

Unfortunately, the facial nerve is the most frequently affected cranial nerve in head and neck trauma. The lesion of the facial nerve are usually

postoperative (removal of cerebellopontine angle tumors, acoustic neuroma surgery, parotid resections for malignancy) or consequences of traffic accidents (brainstem hemorrhage, temporal bone fractures, or lacerations of the face).

To date all modern surgical approaches for treatment of the facial palsy are aimed at creating favourable conditions for accelerated centrifugal regrowth of the facial axons towards their peripheral targets. These include the immediate end-to-end suture of the transected facial nerve (facial-facial anastomosis), suture of the proximal stump of a transected hypoglossal nerve to the distal stump of the facial nerve (hypoglossal-facial anastomosis) and microsurgical implanting of grafts harvested from the sural (2) and/or radial (3) nerve.

Synchronously with the advancing sophisticated neurological techniques and high-tech tools, much attention was given to improve the process of regeneration of the facial motoneurons in the brainstem. Before reaching this goal, however, voluminous experimental work was performed on the reactions in the facial nucleus after injury of the peripheral nerve. The results obtained attempted to answer several major questions: (i) what are the morphological changes that occur in the facial nucleus after transection of the facial nerve, (ii) is there a correlation between the severity of the lesion and the final

neurological outcome, (iii) is the subsequent function as good as normal, and (iv) can the process of regeneration be accelerated?

The aim of the present paper is to provide an overview and interpretation of the answers to these questions, i.e. this review deals with the neuronal and glial changes in the facial nucleus in response to the most common experimental injuries on the extratemporal portion of the facial nerve in adult rats. It includes neither a description of the post-lesion changes along the peripheral nerve fibers themselves, nor any data about the post-traumatic reinnervation of the motor end-plates of the mimetic musculature. Hence, this script is concerned only with the events in the motoneurons and surrounding neuroglia after the peripheral (axonal) lesion of the VII cranial nerve.

CHANGES IN THE FACIAL NUCLEUS AFTER TRANSECTIONS OF THE FACIAL NERVE

• Neuronal and glial changes at the cellular level

• *Neuronal changes*

The first systematic review on the consecutive changes in the facial motoneurons induced by avulsion of the facial nerve in rabbits has been presented by Franz Nissl on 9th November 1890 (4). He demonstrated a disintegration of the "chromatin" bodies in the lesioned perikarya beginning within the first 24 h after the lesion. Later, it became clear that the classical term "chromatolysis" involves the dispersion and hypertrophy of the neuronal rough endoplasmic reticulum (5). However, a recent quantitative analysis of chromatolysis revealed that this reaction starts within 8 h after axon transection (6).

Within 3 days after axon injury, the facial motoneurons increase in profile area (7). The increase of neuronal soma, nuclei, and nucleoli has been measured morphometrically to reach 30-40% between 10 and 21 days post axotomy (8) and has been interpreted as a correlate of the accelerated synthesis of rRNA (9) or as a reflection of the transient transcription of mRNA (8).

Glial changes

The microglial cells proliferate and reach a perineuronal position, thus isolating the neurons from their synaptic inputs (10-13). This displacement of boutons by microglial cells is probably preceded by a loosening of the synaptic contacts, due to some changes in the membranes of the lesioned motoneurons (10).

Neuronal and glial changes at the molecular level

Neuronal changes

The *direct self-defence mechanisms* of the lesioned neurons are immediately activated. One of them is the recently shown expression of nitric oxide synthase (NOS) in axotomized facial motoneurons, which is considered to counteract the massive increase in free radicals consequent to axonal injury and thus supports the anabolic programme for regeneration of the damaged axons (14).

Another molecular mechanism for direct neuronal self-protection may be the migration of the cytosolic enzyme neuron-specific enolase (NSE) into the nuclei of the axotomized neurons (15). The neuronotrophic effect of the intranuclear NSE may be attributed to the resulting pyruvate which has a potent neuronal survival-promoting action by protection of the whole intracellular machinery against peroxide-induced damage (16). A strong support to this hypothesis is the statement that NSE directly promotes the survival of embryonic rat neurons in primary culture (17).

A third molecular protection is the dramatical increase in content of the possibly neurotrophic factor calcitonin gene-related peptide (CGRP) following transection of the facial nerve (18-21). It is known that CGRP stimulates the activity of adenylate cyclase leading to an accumulation of cAMP, thereby regulating acetylcholine receptor synthesis (22,23). On the other hand, CGRP has been suggested a putative diffusible signalling factor for the activation of the surrounding glia (21). This notion is partially confirmed by the established dependence

of glial fibrillary acidic protein (GFAP) mRNA synthesis on high cAMP levels (24). Further, it is suggested that (i) due to the rapid accumulation of interleukin (IL)-6 mRNA in the facial nucleus, IL-6 might also act as an early activating signal for glial cells after axotomy of the facial nerve, and (ii) the triple increase of transforming growth factor (TGF)- β 1 mRNA in the activated microglial cells around regenerating motoneurons provides a long-lasting negative feedback signal controlling glial activation (25).

Also at the mRNA level it has been shown that a particular family of genes might play a crucial role in adaptive plasticity and in long-term changes in the nervous system. These are the immediate early genes (IEG) which transcription is activated rapidly within minutes after axotomy. Damage to the facial axons causes a unique increase in the mRNAs of c-jun and jun B in the facial nucleus (26-29). The clear temporal and spatial correlation between c-jun/jun B mRNA expression and the induction of the CGRP gene in the injured facial nucleus shows that the CGRP gene could be one of the targets of c-jun and jun B (26).

The neuronal survival programme of the axotomized neurons disposes with a wide spectrum of *indirect self-defence mechanisms*, i.e. reactions which are not directly aimed against neuronal death. One of them is the switch from a mode of homeostatic cell maintenance to a mode of intense biosynthetic activity, including both increase of RNA content and uptake of amino acids in motoneurons (5). The activity of ornithine decarboxylase, a key enzyme in the polyamine biosynthesis (responsible for the production of neuronal growth factors), reaches 300% over control (30). The activity of trans-glutaminase, the putable enzyme through which the polyamines exert their effects, also increases (31). The transection of the facial nerve is followed by an increased intraneuronal uptake of glucose (32) and iron, the latter being accompanied by an enhanced immunoreactivity of transferrin receptors (33). The increased consumption of glucose is directed to the pentose phosphate shunt which is also activated after axotomy (7,34,35). The pentose phosphate shunt is known to be involved in the production of

ribose and NADPH. Ribose is necessary for the increased synthesis of RNA, and NADPH furnishes proton equivalents for the synthesis of lipids necessary for membrane restoration during axonal regrowth and sprouting (7). In frame of this intensive regeneration programme, the synthesis of cytoskeletal proteins, tubulin and actin, is also increased (36-38) and newly synthesized "growth associated proteins" (GAP) appear (39).

In addition, studies have also shown that some proteins generated during the axon reaction are not normally found in the perikaryal cytoplasm of adult neurons (40). The developmentally regulated tubulin mRNA and also GAP-43 mRNA have been found to be rapidly induced in regenerating adult facial nuclei (37,41).

There occurs a decrease in the synthesis of transmitter-related compounds, i.e. enzymes or receptor proteins. The activities of some enzymes involved in transmitter metabolism such, as dopaminehydroxylase, tyrosinehydroxylase, choline-acetyltransferase, and acetylcholinesterase (asymmetric and globular forms) decrease in response to axotomy (42-46). The general neuronal activity, as judged upon by the established decrease in activity of cytochrome oxidase (45), is also diminished. In addition, the synthesis of the 68 kD and 150 kD neurofilament proteins is decreased (36).

Glial changes

The retrograde changes at the molecular level after axotomy are not limited only to the neurons, but also involve the surrounding microenvironment of glial cells. By supporting neuronal growth and metabolism glial cells may determine, in part, the degree of recovery after injury. Whereas it has been shown that astroglia actually supports neuronal growth and survival (47-49), cell culture studies indicate that microglia can release cytotoxic agents (50-52). The ramified quiescent microglia comprise a regularly spaced network of cells within the facial nucleus of the rat. In response to sublethal injury of the facial motoneurons (eg. transection of the facial nerve) these resting microglial cells show a dramatic increase in mitotic activity and rapidly migrate toward

the neurons (12,53,54). Upon reaching the neuronal cell surface, the microglial cells displace the afferent synaptic terminals (10) and increase the synthesis of cell adhesion molecules (55) which enable them to bind into uninterrupted rings around the lesioned neurons.

The presence of activated microglial cells around lesioned neurons (regardless of the type of injury), however, does not always result in neuronal death (56,57). Light microscopical and ultrastructural histochemistry revealed high activity of the ectoenzyme 5'-nucleotidase on the membranes of the microglia (58-60). Recently, this finding was further precized by showing that 5'-nucleotidase immunoreactivity is predominantly associated with perineuronal microglia (61) which have been implicated in the lesion-induced detachment and displacement, or "stripping" of synapses from the lesioned motoneurons (10,13,53,61). This enzyme produces some nucleotides and adenosine which are potent regulators of vascular tone, and also affect neural and immune cellular activities (62-66).

A challenging issue for understanding any degenerative CNS disease, especially immune-mediated injury, is whether brain-associated immune cells are capable of doing both phagocytosis and antigen presentation (13,67-79), and their interactions with T-lymphocytes passing through the blood-brain barrier (80-82).

The astroglia response to axon reaction is initially confined to a slight hypertrophy which is difficult to quantify and specify (83). It is the paper of Graeber and Kreutzberg (84) that marks the beginning of a reliable assessment of the participation of astrocytes in the axon reaction, i.e. the immunohistochemical demonstration of GFAP-positive staining in the reactive astrocytes. GFAP is the cell-specific intermediate filament protein in astrocytes and the upregulation of its synthesis is considered a definite feature of activated astroglia. Employing this reliable method, Graeber and Kreutzberg (85) show that three weeks post axotomy the reactive astrocytes start to form thin lamellar processes which separate the neuronal perikarya and adjacent presynaptic boutons for several months. This gross

alignment of postsynaptic membrane and presynaptic boutons is considered a prerequisite for functional recovery (57). The upregulation of GFAP content in the astrocytes starts 12 h after nerve lesioning and the time course and extent of the GFAP synthesis are strongly influenced by successful or unsuccessful regeneration (86). Recent data, however, provide evidence that the increased synthesis of GFAP is preceded by an enhanced coupling capacity of astrocytes: as early as 45-90 min after transection of the facial nerve in rats, there occurs an increase in the immunoreactivity of the predominant gap junction protein in astrocytes connexin 43 (87).

CORRELATION BETWEEN THE SEVERITY OF THE LESION AND THE FINAL NEUROLOGICAL OUTCOME

H Depending on the outcome there are two major groups of experimental injuries. Those from the first group provide conditions that favour neuronal regeneration. Alternatively, the experimental lesion included in the second group impede the axonal resprouting and thus provide models for studying neuronal degeneration and neuronal death.

Following nerve lesions from the *first group* all facial motoneurons resprout branching axons which succeed to reach peripheral targets. Due to the reestablished nerve-muscle contract the motoneuronal perikarya in the brainstem of adult rats are able to survive the insult from the injury, i.e. they successfully regenerate. This "regeneration group" includes the following experimental injuries: crush (8,45,88,89), transection (14,44,46,54,83,87-90), sequential double transection known also as conditioning lesion (7), transection and immediate end-to-end suture (6,56,92,93), transection and immediate suture to the hypoglossal nerve, i.e. hypoglossal-facial anastomosis (HFA)(15,57,94), the latter deserving a special attention.

First, HFA is a standard method for the surgical treatment of facial palsy in man: the end-to-end suture of the freshly transected normal hypoglossal nerve to the distal stump of the transected facial nerve enables the outgrowth of hypoglossal axons into the paralyzed facial plexus. As a result, the

mimetic musculature of the human face is reinnervated up to spontaneous, i.e. subconscious smiling, which indicates a remarkable degree of neuronal plasticity in the brainstem or higher motoric centers of man (95-101). Our own experiments show that despite the resprouting of hypoglossal axonal branches into the mimetic musculature, axons of the transected facial nerve (left untreated during operation) also resprout and reach the facial muscles. Eight weeks after HFA there are 43% more motoneurons (hypoglossal and facial together) that project to the rat whiskerpad muscles than in normal animals. This "hyperinnervation", i.e. the projection of more motoneurons into the target muscle than under normal conditions, is persistent and constant up to 32 weeks post operation (94).

A comparison of the time course of this dual resprouting shows that between 14 and 28 days post operation (dpo) the facial muscles are reached and consequently innervated by hypoglossal motoneurons. This may result in normalization of both trophic secretion and expression of neural cell adhesion molecules (N-CAM) (102) and thus it should prevent any later ingrowth of axons. Furthermore, between 42 and 56 dpo, the total amount of nerve fibers that supply the whiskerpad (facial and hypoglossal together) equals that under normal conditions. Despite of this, both hypoglossal and facial motoneurons continue to send axons to their new or original area of innervation, causing the hyperinnervation. What are the regulatory mechanisms of this development? Some biophysical properties of the normal hypoglossal motoneurons of the rat (103) are almost identical with those of the normal facial neurons of the rat (104,105). This is, however, not the case as the active biophysical characteristics of the hypoglossal and facial motoneurons are concerned. The frequency of discharge of the hypoglossal motoneurons is 5.2 ± 0.6 per minute (106,107), i.e. about 0.1 Hz. This frequency of impulses, however, is far lower than that of the original facial supply of the whiskerpad. During exploration the rats move their mystacial vibrissae back and forth at a rate of about 7 Hz and additionally display a tremorlike movement of the vibrissae at 9 Hz (108). Therefore, the muscles of the whiskerpad are not adequately stimulated by the hypoglossal motor activ-

ity and hence continue the secretion of reinnervation-promoting factor(s) (109).

These new results from the HFA paradigm confirm the major role and highest demands of peripheral targets in the process of reinnervation. Evidently, under this experimental protocol, it seems rather easy to achieve conditions for axonal resprouting and neuronal regeneration in rats. Practically all neurons survive after facial-facial anastomosis (56). Unfortunately, in a vast portion of patients the facial palsy is due to postoperative removal of cerebellopontine angle tumors or acoustic neuroma surgery (2,110) after which there are no regenerating neurons left in the brainstem. Thereanimation of the facial musculature in rats obviously requires the employment of such experimental approaches which prevent the spontaneous axonal regrowth and thus induce neuronal degeneration.

These approaches are the object of *thesecond group* of nerve lesion after which there occurs no resprouting of the facial motoneurons and a large portion of neurons gradually die. The lesions included in this "degeneration group" are: transection and avulsion by gentle traction (88,89), injection of toxic ricin (75,78,111), resection, i.e. transection and removal of 10 mm nerve length (37,56,57,86,94, 112-115).

The most intriguing reaction in the facial nucleus after all severe lesions to the facial motoneurons concerns the microglia. Following any lethal injury of motoneurons the activated microglia do not only express all reviewed immune-related changes (see above), but develop into full-blown macrophages (111). These phagocytic microglia, however, do not only remove the debris of the dead motoneurons, but can present antigens to T-lymphocytes and initiate an immune response (11,78,82). Long-term studies from our laboratory employing retrograde labeling of the facial motoneurons with Fluoro-Gold prior to resection of the facial nerve show that following completion of phagocytic activity, the neuronophages migrate toward adjacent degenerating cells from the same lesioned neuronal pool (112,113,116,117).

IS THE SUBSEQUENT RESERVATION AS GOOD AS NORMAL?

3 Functional recovery after lesions of the facial nerve is usually poor, the major reason for this being the misguidance of the regenerating fibers to inappropriate peripheral targets (92-94,118). Despite the use of presently available microsurgical techniques for repair of injured peripheral nerves, i.e. clean wound, gentle tissue handling, good adaptation and coaptation, minimal number of sutures, and absence of tension (119), there always occurs a substantial mismatching of motoneurons and muscles following transection and subsequent regeneration within a nerve trunk.

After nerve transection in adult animals virtually all neurons survive, but the regenerating axons seem to grow in a relatively random manner resulting in a considerable disarray of the facial nucleus. This loss of somatotopic organization in the facial nucleus following injury of the peripheral nerve is the morphological correlate of the phenomenon of misdirected resprouting termed also excessive reinnervation, aberrant reinnervation, aberrant regeneration, or misdirected regrowth of axons. As a consequence, the coordinated activity of individual muscles is impaired (120-122) and abnormal associated movements (synkinesis), hemifacial spasms, or contractures may develop (123).

IMPROVEMENT OF NEUROMAL REGENERATION

- In an attempt to improve axonal resprouting we performed a pharmacological experiment that was aimed at testing a favourable effect of the calcium channel blocker nimodipine on neuronal plasticity (124-126). Following HFA, nimodipine treatment, and injection of horseradish peroxidase (HRP) into the mimetic musculature of rats, we counted the retrogradely labelled motoneurons in the brainstem at various postoperative survival times. The comparison of these neuron numbers with data from experimental animals not subjected to nimodipine treatment (placebo), showed that the peroral administration of the drug has two beneficial effects (127). First, it shortens the time for the successful sprouting of the axotomized hypoglossal

axons into the facial periphery by 50%. Our quantitative estimates show that in nimodipine-treated animals the sectioned hypoglossal axons reach the whiskerpad muscles 14 days earlier than in placebo-fed rats (first HRP-labelled neurons at 28 dpo). In this way, the proposed favourable effect of nimodipine on neuronal recovery (128,129) is expressed by a significant acceleration of the axonal resprouting. We cannot provide evidence about the morphological and molecular events that accompanied this increased rate of recovery since the precise mechanism of action of the drug is still unknown (130 for review). Anyway, a dual effect on both perikarya and neurites may occur. First, direct neuronotrophic effect has been already suggested: as nimodipine passes the blood-brain barrier and binds to specific dihydropyridine receptors, it possibly prevents the influx of Ca^{2+} into the injured neuronal cell bodies (91,131-135). Second, peripheral beneficial effect of nimodipine on the axonal branches may also involve the fine regulation of intracellular calcium in outgrowing sprouts (136).

These two possible effects of the drug are added to a third factor, i.e. the enhanced hypoglossal sprouting due to the crossed nerve suture: the significant difference in fiber numbers between the facial and the hypoglossal nerve creates a situation in which the hypoglossal axons are guided by 40% more rows of Schwann cells during their regrowth (57,94). In our opinion, it is not the single action, but the combination of all three neuronprotective effects that ensures this improved recovery (126,137).

The second favourable effect of the administration of nimodipine in rats with HFA is that it completely prevents the pathological hyperinnervation and causes a statistically significant suppression of the spontaneous misdirected resprouting of the facial motoneurons. Like in the placebo-fed animals after HFA, a misdirected resprouting of the facial axons occurs (first detected 6 weeks after HFA), but the number of resprouted facial motoneurons remains 38% less than in placebo-fed animals.

Apart from the fact that this lower neuron number does not lead to a hyperinnervation, there seems to occur a strange effect of the drug: on one side, it has

a stimulatory influence on the sutured stump of the transected hypoglossal nerve, and on the other, a suppressive effect on the proximal stump of the transected facial nerve.

Based on two major considerations (i) it is very unlikely that nimodipine may have had a selective action on these two types of brainstem motoneurons, and (ii) the suppression of the facial resprouting starts from its very beginning, we suggest that this strongly limited sprouting of the facial axons is not due to a suppressive effect of nimodipine, but rather to the very rapidly occurred hypoglossal reinnervation of the whiskerpad. In placebo-fed animals, the first axons of the facial motoneurons reach whiskerpad muscles between 28 and 42 dpo in a period when already about 570 hypoglossal motoneurons project there. This retardation (within the next 2 weeks) and the ratio between hypoglossal and facial neurons of 2:1 provide evidently good grounds for a competitive relationship which yields in a prolonged hyperinnervation of the whiskerpad (94). Under nimodipine treatment, however, the situation is very different. Due to accelerated resprouting (see above), the hypoglossal axons reach the facial periphery very early and are the only neurites in the target area for a twice as long period (from the 2nd till the 6th postoperative week) than in placebo-fed animals.

Till 7 dpo, no motor axons supply the mimetic musculature and the denervated facial muscles intensively secrete molecules that promote neuronal survival (109) and accumulate N-CAM that enhance their attractiveness to axons (102). Between 7 and 14 dpo the facial muscles are reached and innervated by the hypoglossal motoneurons. This reinnervation is, however, completely different from the one that occurs in the placebo-fed animals.

During reinnervation following nerve crush, there is a period when the motoneurons reach neuromuscular junctions, but do not release neurotransmitters (138,139). The first miniature end-plate potentials (MEPP) reappear approximately 2 weeks after the crush-lesion (140). Especially in these early stages of reinnervation, however, the Ca^{2+} -dependent release mechanisms are highly efficient, i.e. the influx of Ca^{2+} increases the frequency of MEPP. The

reason for this is "the low buffering capacity for free intracellular Ca^{2+} of the regenerating nerve terminals, which would allow a stronger stimulatory action of the Ca^{2+} which has entered the presynaptic ending". In spite of this very high efficiency, however, the MEPP frequency remains far below normal (141).

The nimodipine treatment reduces the calcium influx into the axoplasm of the resprouting nerve fiber (136) and further reduces its buffering capacity for Ca^{2+} . This might render the responsiveness of the terminals to Ca^{2+} even stronger which would yield MEPP with even higher frequency. In this way, a much earlier and qualitatively better reinnervation of the facial muscles is achieved, that leads to a partial normalization both of trophic secretion and N-CAM expression. This results in a significant decrease of the misdirected re-sprouting of facial nerve fibers, there occurs no-competitive relationship between the facial and hypoglossal motoneurons, and no hyperinnervation.

REFERENCES

1. Bento RF, Miniti AM. Anastomosis of the intratemporal facial nerve using fibrin tissue adhesive. *Ear Nose Throat J* 1993; 72: 663-672
2. Braam MJI, Nicolai JPA. Axonal regeneration rate through cross-face nerve grafts. *Micro-surgery* 1993; 14: 589-591
3. Vaughan ED, Richardson D. Facial nerve reconstruction following ablative parotid surgery. *Br J Oral Maxillofac Surg* 1993; 31: 274-280
4. Nissl F. liber die Veränderungen der Ganglienzellen am Facialiskern des Kaninchens nach Ausreissung der Nerven. *Allg Z Psychiat* 1891; 48: 197-198
5. Lieberman AR. The axon reaction: a review of the principal features of the perikaryal response to axon injury. *Int Rev Neurobiol* 1971; 14: 49-125
6. Neiss WF, Schulte E, Guntinas-Lichius O, Gunkel A, Stennert E. Quantification of chromatolysis in motoneurons of the Wistar rat. *Ann Anat (Suppl)* 1993; 175: 6

7. Tetzlaff W, Kreutzberg GW. Enzyme changes in the rat facial nucleus following a conditioning lesion. *Exp Neurol* 1984; 85: 547-564
8. Vaughan DW. Effects of advancing age on the central response of rat facial neurons to axotomy: Light microscope morphometry. *AnatRec* 1990; 228:211-219
9. Jones KJ, La Velle A. Differential effects of axotomy on immature and mature hamster facial neurons: a time course study of initial nucleolar and nuclear changes. *J Neurocytol* 1986; 15: 197-206
10. Blinzinger K, Kreutzberg GW. Displacement of synaptic terminals from regenerating motoneurons by microglial cells. *Z Zellforsch* 1986; 85: 145-157
11. Graeber MB, Streit WJ, Kreutzberg GW. Formation of microglia-derived macrophages is blocked by adriamycin. *Acta Neuropathol* 1989; 78: 348-358
12. Kreutzberg GW. Uber perineuronale Mikrogliazellen (Autoradiographische Untersuchungen). *Acta Neuropathol (Suppl IV)* 1968; 141-145
13. Streit WJ, Graeber MB, Kreutzberg GW. Functional plasticity of microglia: a review. *Glia* 1988; 1: 301-307
14. Yu WHA. Nitric oxide synthase in motor neurons after axotomy. *J HistochemCytochem* 1994; 42: 451-457
15. Angelov DN, Neiss WF, Gunkel A, Guntinas-Lichius O, Stennert E. Axotomy induces intranuclear immunolocalization of neuron-specific enolase (NSE) in facial and hypoglossal neurons of the rat. *J Neurocytol* 1994; 23: 218-233
16. Perez-Polo JR, Foreman PJ, Jackson GR, Shan DE, Tagliatela G, Thorpe LW, Werrbach-Perez K. Nerve growth factor and neuronal cell death. *Mol Neurobiol* 1990; 4: 57-91
17. Takei N, Kondo J, Nagaike K, Ohsawa K, Kato K, Kohsaka S. Neuronal survival factor from bovine brain is identical to neuron-specific enolase. *J Neurochem* 1991; 57: 1178-1184
18. Arvidsson U, Johnson H, Piehl H, Cullheim S, Hokfelt T, Risling M, Terenius L, Ulfhake B. Peripheral nerve section induces increased levels of calcitonin gene-related peptide (CGRP)-like immunoreactivity in axotomized motoneurons. *Exp Brain Res* 1990; 79: 212-216
19. Haas CA, Streit WJ, Kreutzberg GW. Rat facial motoneurons express increased levels of calcitonin gene-related peptide mRNA in response to axotomy. *J Neurosci Res* 1990; 27: 270-275
20. Moore RY. Cranial motoneurons contain either galanin- or calcitonin gene-related peptide like immunoreactivity. *J Comp Neurol* 1989; 282: 512-522
21. Streit WJ, Dumoulin FL, Raivich G, Kreutzberg GW. Calcitonin gene-related peptide increases in facial motoneurons after peripheral nerve transection. *Neurosci Lett* 1989; 101: 143-148
22. Laufer R, Changeux JP. Calcitonin gene-related peptide elevates cyclic AMP levels in chick skeletal muscle: possible neurotrophic role for a coexisting neuronal messenger. *EMBO J* 1987; 6: 901-906
23. New HV, Mudge AW. Calcitonin gene-related peptide regulated muscle acetylcholine receptor synthesis. *Nature* 1986; 323: 809-811
24. Shafit-Zagardo B, Kume-Iwaki A, Goldman JE. Astrocytes regulate GFAP mRNA levels by cyclic AMP and protein kinase C-dependent mechanisms. *Glia* 1988; 1: 721-745
25. Kiefer R, Lindholm D, Kreutzberg GW. Interleukin-6 and transforming growth factor- β mRNAs are induced in rat facial nucleus following motoneuron axotomy. *Eur J Neurosci* 1993; 5: 775-781,
26. Haas CA, Donath C, Kreutzberg GW. Differential

- expression of immediate early genes after transection of the facial nerve. *Neuroscience* 1993; 53: 91-99
27. Herdegen T, Kovary K, Leah J, Bravo R. Specific temporal and spatial distribution of JUN, FOS, and KROX-24 proteins in spinal neurons following noxious transsynaptic stimulation. *J Comp Neurol* 1991; 313: 178-191
 28. Herdegen T, Kummer W, Fiallos CE, Leah J, Bravo R. Expression of c- JUN, JUN B and JUN D proteins in rat nervous system following transection of vagus nerve and cervical sympathetic trunk. *Neuroscience* 1991; 45: 413-422
 29. Herdegen T, Tölle T, Bravo R, Zieglgansberger W, Zimmermann M. Sequential expression of JUN B, JUN D and FOS B proteins in rat spinal neurons: cascade of transcriptional operations during nociception. *Neurosci Lett* 1991; 129: 221 -224
 30. Tetzlaff W, Kreutzberg GW. Ornithine decarboxylase in motoneurons during regeneration. *Exp Neurol* 1985; 89: 679-688
 31. Tetzlaff W, Gilad VH, Leonard C, Bisby MA, Gilad GM. Retrograde changes intransglutaminase activity after peripheral nerve injuries. *Brain Res* 1988; 445: 142-146
 32. Kreutzberg GW, Emmert H. Glucose utilization of motor nuclei during regeneration: a (¹⁴C) 2-deoxyglucose study. *Exp Neurol* 1980; 70: 712-716
 33. Graeber MB, Raivich G, Kreutzberg GW. Increased transferring receptor expression by regenerating facial motoneurons. *Soc Neurosci* 1988; 14: 1055A
 34. Harkönen MHA, Kauffman FC. Metabolic alterations in the axotomized superior cervical ganglion of the rat. II. The pentose phosphate pathway. *Brain Res* 1974; 65: 141-157
 35. Kreutzberg GW. Changes of coenzyme (TPN) diaphorase and TPN-linked dehydrogenase during axonal reaction in the nerve cell. *Nature* 1963; 199: 393-394
 36. Tetzlaff W, Bisby MA, Kreutzberg GW. Changes in cytoskeletal proteins in the rat facial nucleus following axotomy. *J Neurosci* 1988; 8: 3181-3189
 37. Tetzlaff W, Alexander SW, Miller FD, Bisby MA. Response of facial and rubrospinal neurons to axotomy: changes in mRNA expression for cytoskeletal proteins and GAP-43. *J Neurosci* 1991; 11: 2528-2544
 38. Bisby MA, Tetzlaff W. Changes in cytoskeletal protein synthesis following axon injury and during regeneration. *Mol Neurobiol* 1992; 6: 107-123
 39. Willard M, Skene JHP. Molecular events in axonal regeneration. In: Nicholls J G, editor. *Repair and Regeneration of the Nervous System*, Springer, Berlin 1982; 71
 40. Griffith A, La Veile A. Developmental protein changes in normal and chromatolytic facial nerve nuclear regions. *Exp Neurol* 1971; 33: 360-371
 41. Miller FD, Tetzlaff W, Bisby MA, Fawcett JW, Milner RJ. Rapid induction of the major embryonic alpha-tubulin mRNA, T alpha 1, during nerve regeneration in adult rats. *J Neurosci* 1989; 9: 1452-1463
 42. Engel AK, Kreutzberg GW. Changes in acetylcholinesterase molecular forms in regenerating motor neurons. *Neuroscience* 1986; 18: 467-473
 43. Engel AK, Tetzlaff W, Kreutzberg GW. Axonal transport of 16S acetylcholinesterase is increased in regenerating peripheral nerve in guinea-pig, but not in the rat. *Neuroscience* 1988; 24 :729-738
 44. Hoover DB, Hancock JC. Effect of facial nerve transection on acetylcholinesterase and (³H) quinuclidinyl benzilate binding in rat facial nuclei. *Neuroscience* 1985; 15: 481-487
 45. Vaughan DW. The effects of age on enzyme activities in rat facial nucleus following axotomy: acetylcholinesterase and cytochrome oxidase. *Exp Neurol* 1990; 109: 224-236
 46. Senda E, Simmons DM, Wada E, Wada K,

- Swanson E. RNA levels of neuronal nicotinic acetylcholine receptor subunits are differentially regulated in axotomized facial motoneurons: an in situ hybridization study. *Mol Brain Res* 1990; 8: 349-353
47. Manthorpe M, Rudge JS, Varon S. Astroglial cell contributions to neuronal survival and neuritic outgrowth. In: Federoff S, Vernadakis A, editors. *Astrocytes*. Academic Press, New York 1986; 2: 315-376
48. Manthorpe M, Ray J, Pettman B, Varon S. Giliary neurotrophic factors. In: Rush A, editor. *Nerve Growth Factors*. Wiley, New York 1989; 31-36
49. Henderson ChE, Phillips HS, Pollock RA, Davies AM, Lemeulle C, Armanini M, Simpson LC, Moffet B, Vandlen RA, Koliatsos VE, Rosenthal A. GDNF: A potent survival factor for motoneurons present in peripheral nerve and muscle. *Science* 1994; 266: 1062-1064
50. Colton CA, Gilbert DL. Production of superoxide anions by a CNS macrophage, the microglia. *FEES Lett* 1987; 223: 284-288
51. Guilian D, Baker TJ. Characterization of amoeboid microglia isolated from developing mammalian brain. *J Neurosci* 1986; 6: 2163-2178
52. Guilian D, Vaca K, Corpuz M. Brain glia release factors with opposing actions upon neuronal survival. *J Neurosci* 1993; 13: 29-37
53. Graeber MB, Tetzlaff W, Streit WJ, Kreutzberg GW. Microglial cells but not astrocytes undergo mitosis following rat facial nerve axotomy. *Neurosci Lett* 1988; 85: 317-321
54. Kreutzberg GW. Autoradiographische Untersuchungen über die Beteiligung von Gliazellen an der axonalen Reaktion im Facialiskern der Ratte. *Acta Neuropathol* 1966; 17: 149-161
55. Moneta M, Gehrman J, Topper R, Banati RB, Kreutzberg GW. Cell adhesion molecule expression in the regenerating facial nucleus. *J Neuroimmunol* 1993; 45: 203-206
56. Guntinas-Lichius O, Neiss WF, Gunkel A, Stennert E. Nerve suture or resection of the facial nerve: Differences in microglial, astroglial, synaptic and motoneuron responses in the facial nucleus of the rat brainstem. *Europ Arch Otorhinolaryngol* 1994; In press
57. Neiss WF, Schulte E, Guntinas-Lichius O, Angelov DN, Gunkel A, Stennert E. The hypoglossal-facial anastomosis as model of neuronal plasticity in the rat. *Ann Anat* 1992; 174: 419-433
58. Kreutzberg GW, Barren KD. 5'-Nucleotidase of microglial cells in the facial nucleus during axonal reaction. *J Neurocytol* 1978; 17: 601-610
59. Kreutzberg GW. The motoneuron and its micro-environment responding to axotomy. In: Das GD, Wallace RB, editors. *Neural Transplantation and Regeneration*. Springer, New York 1985; 271-276
60. Kreutzberg GW, Heymann D, Reddington M. 5'-Nucleotidase in the nervous system. In: Kreutzberg GW, Redington M, Zimmermann H, editors. *Cellular Biology of Ectoenzymes*, Springer. New York 1986; 147-164
61. Schoen SW, Graeber MB, Kreutzberg GW. 5'-Nucleotidase immunoreactivity of perineuronal microglia responding to rat facial nerve axotomy. *Glia* 1992; 6: 314-317
62. Dunwiddie TV, Hoffer BJ. Adenine nucleotides and synaptic transmission in the in vitro rat hippocampus. *Br J Pharmacol* 1980; 69: 59-68
63. Lee KS, Schubert P, Heinemann U. The anticonvulsive action of adenosine: a postsynaptic dendritic action by a possible endogenous anticonvulsant. *Brain Res* 1984; 321: 160-164
64. DiVirgilio F, Pizzo P, Zanovello P, Bronte V, Collavo D. Extracellular ATP as a possible mediator of cell mediated cytotoxicity. *Immunol*

- Today 1990; 11:274-277
65. Hassessian HM. Old, new and not yet exploited purinergic vasomechanisms of the pulmonary circulation, *Biomed Rev* 1994; 3: 11-25
 66. Linden J. Cloned adenosine A3 receptors: pharmacological properties, species differences and receptor functions. *Trends Pharmacol Sci* 1994; 15: 298-306
 67. Graeber MB, Streit WJ, Kreutzberg GW. Axotomy of rat facial nerve leads to increased CR3 complement receptor expression by activated microglial cells. *J Neurosci Res* 1988; 21: 18-24
 68. Milligan CE, Cunningham TJ, Levitt P. Brain macrophages transiently target certain axon tracts. *Soc Neurosci* 1989; 15: 445A
 69. Milligan CE, Levitt P, Cunningham TJ. Brain macrophages and microglia respond differently to lesion of the developing and adult visual system. *J Comp Neurol* 1991; 314: 136-146
 70. Ogawa M, Araki M, Naito M, Takeya M, Takahashi K, Yoshida M. Early changes of macrophage-like immunoreactivity in the rat inferior olive after intraperitoneal 3-acetylpyridine injection. *Brain Res* 1993; 610: 135-140
 71. Streit WJ, Graeber MB. Heterogeneity of microglial and perivascular cell populations: Insights gained from the facial nucleus paradigm. *Glia* 1993; 7: 68-74
 72. Perry VH, Hume DA, Gordon S. Immunohistochemical localization of macrophages and microglia in the adult and developing mouse brain. *Neuroscience* 1985; 15: 313-326
 73. Rinaman L, Milligan CE, Levitt O. Persistence of Fluoro-Gold following degeneration of labelled motoneurons is due to phagocytosis by microglia and macrophages. *Neuroscience* 1991; 44: 765-776
 74. Akiyama H, McGeer PL. Microglial response to 6-hydroxydopamine-induced substantia nigra lesions. *Brain Res* 1989; 489: 247-253
 75. Streit WJ, Graeber MB, Kreutzberg GW. Peripheral nerve lesion produces increased levels of major histocompatibility complex antigens in the central nervous system. *J Neuroimmunol* 1989; 21: 117-123
 76. Hickey WE, Hsu BL, Kimura H. Perivascular microglial cells of the CNS are bone marrow-derived and present antigen *in vivo*. *Science* 1988; 239: 290-292
 77. Maehlen J, Ollson T, Zachau A, Klareskog L, KYistensson K. Local enhancement of major histocompatibility complex (MHC) class I and II expression and cell infiltration in experimental allergic encephalomyelitis around axotomized motor neurons. *J Neuroimmunol* 1989; 23: 125-132
 78. Streit WJ, Graeber MB, Kreutzberg GW. Expression of Ia Antigen on perivascular and microglial cells after sublethal and lethal motor neuron injury. *Exp Neurol* 1989; 105: 115-126
 79. Merrill JE. Tumor necrosis factor alpha, interleukin 1 and related cytokines in brain development: normal and pathological (review). *Dev Neurosci* 1992; 14: 1-10
 80. Hickey WE, Hsu BL, Kimura H. T-lymphocyte entry into the central nervous system. *J Neurosci Res* 1991; 28: 254-260
 81. Wekerle H, Linington C, Lassmann H, Meyernann R. Cellular immune reactivity within the CNS. *Trends Neurosci* 1986; 9: 271-277
 82. Graeber MB. Microglia, macrophages and the blood-brain barrier. In: Kreutzberg GW, editor. *Methods in Microglial Research. Workshop at the 15th Annual Meeting of ENA, Munich, Germany 1992; 12-15*
 83. Cammermeyer J. Astroglial changes during

- retrograde atrophy of nucleus facialis in mice. *J Comp Neurol* 1955; 102: 133-150
84. Graeber MB, Kreutzberg GW. Astrocytes increase in glial fibrillary acidic protein during retrograde changes of facial motor neurons. *J Neurocytol* 1986; 15: 363-373
85. Graeber MB, Kreutzberg GW. Delayed astrocyte reaction following facial nerve axotomy. *J Neurocytol* 1988; 17: 209-220
86. Tetzlaff W. Increased glial fibrillary acidic protein synthesis in astrocytes during retrograde reaction of the rat facial nucleus. *Glia* 1988; 1: 90-95
87. Rohlmann A, Laskawi R, Hofer A, Dermietzel R, Wolff J. Astrocytes as rapid sensors of peripheral axotomy in the facial nucleus of rats. *Neuroreport* 1994; 5: 409-412
88. Soreide AJ. Variations in the axon reaction after different types of nerve lesion. Light and electron microscopic studies on the facial nucleus of the rat. *Acta Anat* 1981; 110: 173-188
89. Soreide AJ. Variations in the perineuronal glial changes after different types of nerve lesion: Light and electron microscopic investigations on the facial nucleus of the rat. *Neuropathol Appl Neurobiol* 1981; 7: 195-204
90. Cammermeyer J. Species differences in acute retrograde neuronal reaction in the facial and hypoglossal nuclei. *J Hirnforsch* 1969; 11: 13-29
91. Van der Kerckhoff W, Drewes LR. Transfer of the Ca-antagonists nifedipine and nimodipine across the blood-brain barrier and their regional distribution *in vitro*. *J Cerebr Blood Flow Metab (Suppl)* 1985; 5: 459-460
92. Aldskogius H, Thomander L. Selective reinnervation of somatotopically appropriate muscles after facial nerve transection and regeneration in the neonatal rat. *Brain Res* 1986; 375: 126-134
93. Tromander L. Reorganization of the facial motor nucleus after peripheral nerve regeneration. *Acta Otolaryngol (Stockh)* 1984; 97: 619-626
94. Angelov DN, Gunkel A, Stennert E, Neiss WF. Recovery of original nerve supply after hypoglossal-facial anastomosis causes permanent motoric hyperinnervation of the whiskerpad muscles in the rat. *J Comp Neurol* 1993; 338: 214-224
95. Hammerschlag PE. Hypoglossal facial nerve anastomosis for correction of facial nerve paralysis following cerebello-pontine angle surgery. In: Ransohoff J, editor. *Modern Techniques in Surgery*. Mount Kisco, New York 1987; 37: 2-12
96. Kreutzberg GW. Acute neuronal reaction to injury. In: Nicholls JG, editor. *Repair and Regeneration of the Nervous System*. Springer Verlag, Berlin 1982; 57-69
97. May M (1986) Surgical rehabilitation of facial palsy. In: May M, editor. *The Facial Nerve: A Total Approach*. Thieme, New York 1986; 695-777
98. May M, Sorol SM, Mester SJ. Hypoglossal-facial nerve interpositional-jump graft for facial reanimation without tongue atrophy. *Otolaryngol Head Neck Surg* 1991; 104: 818-825
99. Miehke A, Stennert E, Arold R, Chilla R, Penzholz H, Kuhner A, Sturrn V, Haubrich J. Chirurgie der Nerven im HNO-Bereich (außer Nn. stato-acusticus and olfactorius). *Arch Oto-Rhino-Laryngol* 1981; 231: 389-449
100. Pitty LF, Tator CH. Hypoglossal-facial nerve anastomosis for facial nerve palsy following surgery for cerebellopontine angle tumors. *J Neurosurg* 1992; 77: 724-731
101. Stennert E. Hypoglossal facial anastomosis: Its significance for modern facial surgery. II. Combined approach in extratemporal facial nerve reconstruction. *Clin Plast Surg* 1979; 6: 471-486

102. Covault J, Sanes JR. Neural cell adhesion molecule (N-C AM) accumulates in denervated and paralyzed skeletal muscles. *Proc Natl Acad Sci USA* 1985; 82: 4544-4548
103. Haddad GG, Donnelly DF, Getting PA. Biophysical properties of hypoglossal neurons in vitro: intracellular studies in adult and neonatal rats. *J Appl Physiol* 1990; 69: 1509-1517
104. Aghajanian G, Rasmussen K. Intracellular studies in the facial nucleus illustrating a simple new method for obtaining viable motoneurons in adult rat brain slices. *Synapse* 1989; 3: 331-338
105. Larkman PM, Penington NJ, Kelly JS. Electrophysiology of adult rat facial motoneurons: the effects of serotonin (5-HT) in a novel in vitro brainstem slice. *J Neurosci Methods* 1989; 28: 133-146
106. Morin D, Hennequin S, Monteau R, Hilaire G. Serotonergic influences on central respiratory activity: an in vitro study in the newborn rat. *Brain Res* 1990; 535: 281-287
107. Morin D, Monteau R, Hilaire G. Compared effects of serotonin on cervical and hypoglossal inspiratory activities: an in vitro study in the new-born rat. *J Physiol* 1992; 451: 605-629
108. Semba K, Egger MD. The facial "motor" nerve of the rat: Control of vibrissal movement and examination of motor and sensory components. *J Comp Neurol* 1986; 247: 144-158
109. Sanes JR. More nerve growth factors? *Nature* 1984; 307: 500
110. King TT, Sparrow OC, Arias JM. Repair of the facial nerve after removal of cerebello-pontine angle tumors: a comparative study. *J Neurosurg* 1993; 78: 720-725
111. Streit WJ, Kreutzberg GW. Response of endogenous glial cells to motor neuron degeneration induced by toxic ricin. *J Comp Neurol* 1988; 268: 248-263
112. Angelov DN, Gunkel A, Schleucher Ch, Krebs C, Stennert E, Neiss WF. Heterogeneity of the microglial/macrophage response in a degenerating motoneuronal cell pool prelabeled with Fluoro-Gold, In: Elsner R, Breer H, editors. *Proceedings of the 22nd Gottingen Neurobiology Conference*. Thieme, Stuttgart 1994; 2: 245
113. Angelov DN, Gunkel A, Stennert E, Neiss WF. Phagocytic microglia during delayed neuronal loss in the facial nucleus of the rat. Time course of the neuronofugal migration of brain macrophages, *Glia* 1995; In press
114. Kreutzberg GW, Tezlaff W, Toth L (1984) Cytochemical changes of cholinesterases in motor neurons during regeneration. In: Brzin M, Kiauta T, Barnard TA, editors. *Cholinesterases: Fundamental and Applied Aspects*, de Gruyter, Berlin 1984; 274-288
115. Saika T, Senda E, Noguchi K, Sato M, Kubo T, Matsunaga T, Tohyama M. Changes in expression of peptides in rat facial motoneurons after facial nerve crushing and resection. *Mol Brain Res* 1991; 11: 187-196
116. Neiss WP, Angelov DN, Gunkel A, Grebs C, Stennert E. Redistribution of Fluoro-Gold from facial motoneurons after long-term axotomy. *Ann Anat (Suppl)* 1994; 176: 69
117. Graeber MB, Streit WJ, Kreutzberg GW. Identity of ED2-positive perivascular cells in rat brain. *J Neurosci Res* 1989; 22: 103-106
118. Matsumoto K. Observation of motoneuron after recovery from experimental facial nerve paralysis. *Nippon Jibinkoka Gakai Kaiho* 1992; 95: 373-380
119. Millesi H. Nerve suture and grafting to restore the extratemporal facial nerve. *Clin Plast Surg* 1979; 6: 331-341
120. Monserrat L, Benito M. Facial synkinesis and

- aberrant regeneration of the facial nerve. *Adv Neurol* 1988; 47: 9-29
121. Sumner AJ. Aberrant reinnervation. *Muscle Nerve* 1990; 13: 801-803
122. Wasserschaff M. Coordination of reinnervated muscle and reorganization of spinal cord motoneurons after nerve transection in mice. *Brain Res* 1990; 515: 241-246
123. Yagi N, Nakatani H. Crocodile tears and thread test of lacrimation. *Ann Otol Rhinol Laryngol (Suppl)* 1986; 95: 13-16
124. Gispén WH, Schuurman T, Traber J. Nimodipine and neural plasticity in the peripheral nervous system of adult and aged rats. In: Morad M, Naylor W, Kazda S, Schramm M, editors. *The Ca²⁺ Channel: Structure, Function and Implication*. Springer, Berlin, Heidelberg, New York, Tokyo 1988; 491-502
125. Schurman T, Klein H, Beneke M, Traber J. Nimodipine and motor deficits in the aged rat. *Neurosci Res Commun* 1987; 1: 9-16
126. Van der Zee CEEM, Brakkee JH, Gispén WH. Putative neurotrophic factors and functional recovery from peripheral nerve damage in the rat. *Br J Pharmacol* 1991; 103: 1041-1046
127. Neiss WF, Angelov DN, Gunkel A, Stennert E. Nimodipine accelerates the sprouting of axotomized motor neurons following hypoglossal-facial anastomosis in the rat. *J Neurotrauma (Suppl)* 1993; 10: 55
128. Nelson C, Finger S, Simons D. Effects of nimodipine on two neurologic measures sensitive to sensorimotor cortex damage. *Exp Neurol* 1993; 119:302-308
129. Poplawsky A. Nimodipine accelerates recovery from the hyperemotionality produced by septal lesions. *Behav Neural Biol* 1990; 53:133-139
130. Wadworth AN, McTavish D. Nimodipine. A review of its pharmacological properties and therapeutic efficacy in cerebral disorders. *Drugs Aging* 1992; 2: 262-286
131. Belleman P, Schade A, Towart R. Dihydropyridine receptors in rat brain labelled with [³H] nimodipine. *Proc Natl Acad Sci USA* 1983; 80: 2356-2360
132. Kazda S, Garthoff B, Luckhaus G. Prevention of acute and chronic cerebrovascular damage with nimodipine in animal experiments. In: Betz E, Deck K, Hoffmeister F, editors. *Nimodipine: Pharmacological and Clinical Properties*. Schatauer, Stuttgart 1985; 31-43
133. Shanne FAX, Kane AB, Young EE, Farber JL. Calcium dependence of toxic cell death: a final common pathway. *Science* 1979; 206: 700-702
134. Siesjö B. Cell damage in the brain: a speculative synthesis. *J Cerebr Blood Flow Metab* 1981; 1: 155-185
135. Simon RP, Griffiths T, Evans MC, Swane JH, Meldrum BS. Calcium overload in selectively vulnerable neurons of the hippocampus during and after ischemia: an electron microscopic study in the rat. *J Cerebr Blood Flow Metab* 1984; 4: 350-361
136. Kater SB, Mattson MP, Cohan C, Conner J. Ca²⁺ regulation of the neural growth cone. *Trends Neurosci* 1988; 11: 315-320
137. Varon S. Factors promoting the growth of the nervous system. *Neuroscience* 1985; 3:62
138. Dennis MJ, Miledi R. Non-transmitting neuromuscular junctions during an early stage of end plate reinnervation. *J Physiol* 1974; 239: 553-570
139. Dennis MJ, Miledi R. Characteristics of transmitter release at regenerating frog neuromuscular junctions. *J Physiol* 1974; 239: 571-594
140. McArdle JJ, Albuquerque XE. A study of the

reinnervation of fast and slow mammalian muscles. *J Gen Physiol* 1973; 61: 1-23

141. Carmignoto G, Finesso M, Siliprandi R, Gorio A. Muscle reinnervation. I. Restoration of transmitter release mechanisms. *Neuroscience* 1983; 8: 393-401

Received for publication 5 September 1994

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