

EDITORIAL

**LEYDIG CELL-IMMUNE CELL INTERACTION: AN EXAMPLE OF
NEUROENDOCRINE-IMMUNE COMMUNICATION IN TESTIS****Kamen P. Valchanov, Anton B. Tonchev, Olawale A.R. Sulaiman, and George N. Chaldakov***Laboratory of Electron Microscopy, Department of Anatomy and Histology, Medical University of Varna, Varna, Bulgaria*

"Never before has the pace of change been so great or so widespread; nothing will ever be the same again. What an opportunity!"

Kathleen L. Wishner

Things will never be the same again.
Diabetes 1996; 45: 541-543

• In her paper "Things will never be the same again" Dr Kathleen L. Wishner (Lilly Corporate Center, Indianapolis, IN) quoted Alvin Toffler's book *Future Shock* written in 1970. Toffler defined "future shock" as a time phenomenon, a product of the greatly accelerated change in society. The scientific research itself is a demonstration of this accelerated change. In particular, data systematized by Davidoff *et al* (1) in this volume of *Biomedical Reviews* indicate the change in the understanding of the nature and origin of Leydig cells of the human testis.

See Review on page 11

Paracrine, autocrine and intracrine mechanisms are obviously important in all organs since they enable the efficient coordination of functions of the different cell types that constitute these organs. The testis needs such mechanisms not only because of the presence of many different cell types, e.g. Leydig cells, Sertoli cells, peritubular myofibroblasts, germ cells, and immune cells, but also because of the changing requirements to coordinate the functions of these cells in a cyclic and time-dependent manner (1,2). Furthermore, all these events have to be coordinated with different body functions. This is accom-

plished by an interaction between paracrine and endocrine signalling, the Leydig cell being a component of the hypothalamic-pituitary-gonadal axis.

Using antibodies directed against a large panel of neuronal and neuroendocrine marker substances, Davidoff *et al* (1) provide conceptual evidence strongly suggestive of a dual, endocrine and neuroendocrine nature and of a neural crest origin of human Leydig cells. Thus these authors propose Leydig cells as a new member of the diffuse neuroendocrine system.

Recently, the bidirectional interaction between the neuroendocrine and immune systems is a subject of intensive research (3-8). The reasons for this are: (I) cytokines as well as neurotrophins and neuropeptides are mediators of both neural and immune responses, (II) neuroendocrine and immune cells have common receptors for hormones, cytokines, neurotrophins, and neurotransmitters, and (III) immune cells produce over 20 different neuroendocrine peptides (5,8) and neurotrophins. Also, cytokines exert pronounced endocrine effects on the hypothalamic-pituitary-adrenocortical axis (6,8) and the hypothalamic-pituitary-gonadal axis (4, 6, 9).

Collectively, these findings (3-8), Davidoff and colleagues' data (1 and Refs therein) and the recent results about Leydig cell-immune cell interactions (9-16) suggest that a local neuroendocrine-immune link may exist within the testis. The Leydig cell-macrophage interaction in rodent testis was described (10-12), suggesting a role of testicular macrophages in both regulation of Leydig cell functions and promotion of Leydig cell development. Yet another part of this testicular neuroendocrine-immune cross-talk is the possible involvement of Leydig cells in estrogen-induced mast cell proliferation in the rat testis, where Leydig cells are proposed to regulate interstitial mast cells proliferation *via* negative paracrine way (13-15). Indeed, Leydig cell express and/or secrete regulatory modules (1, 4, 6, 9), with both activating and inhibitory effects on testicular immune cells. For example, the Leydig cell-derived substance P, vasoactive intestinal polypeptide and (3-endorphin (1) may affect mast cell activity (16, 17) and T cell proliferation (6, 8). Recently we observed mast cell increase in the heart after suppressing the Leydig cells by estrogen treat-

ment of newborn rats indicating that Leydig cells also influence extratesticular immunocytes *via* negative endocrine way (15). Not surprisingly, mast cells and Leydig cells were described to proliferate simultaneously in the testis after selective Leydig cell destruction by ethane dimethane sulfonate (11, see also 1 and Refs 45, 65 therein). Hence, it worths to examine whether mast cell-derived neurotrophins, e.g. nerve growth factor (18) and leukemia inhibitory factor (19), exert some influence on Leydig cells.

Altogether these data would suggest that the Leydig cell-immune cell link is an example of neuroendocrine-immune communication in testis (Fig. 1). We realize that the rapidly developing tale of this interaction warns us that generalization may be premature, and it is better to conclude that there is an *opportunity* here for (*I*) additional data (31), (*I'*) new interpretations of hitherto unexplained, "neurotrophic" facts (32-34, see also 1 and Refs 218, 219 therein), and (*Hi*) broadening the international scientific community involved in this research field

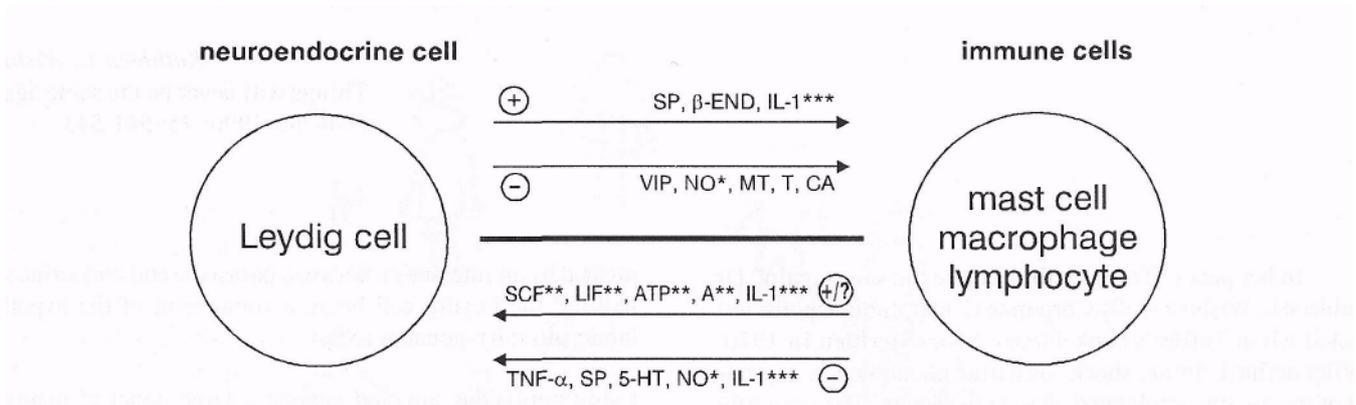


Figure 1. Suggested bidirectional paracrine links between neuroendocrine and immune cells in the testis. A possible cross-talk between Leydig and immune cells via neuroendocrine peptides produced by these cells (1, 5, 8), herein depicted *l*-endorphin (*fi*-END) (16, 17) only, should also be considered.

* effects of nitric oxide (NO) on immune cells (20) may also be depicted in Fig. 11 of Davidoff *et al* (21), illustrating schematically the possible functional significance of Leydig cell-produced NO.

** as found for nerve growth factor receptors (1), we suggest that the receptors for these neurotrophic cytokines (19, 22, 23) as well as some purinoceptors (24, 25) may also be considered in further studies of Leydig cells.

*** note that interleukin-1 (IL-1) stimulates Leydig cell proliferation and inhibits steroidogenesis (6), and induces histamine release from mast cells (26) and neuropeptide Y expression in Leydig cells (1, their Ref 137; see also 1, their Ref 119 for Leydig cell-derived IL-1).

Other abbreviations and respective references: SP - substance P (1, 16, 17, 19), VIP - vasoactive intestinal peptide (6, 16, 17), MT - melatonin (27), T - testosterone (7), CA - catecholamines (28), SCF - stem cell factor (mast cell growth factor, c-kit ligand) (22), LIF - leukemia inhibitory factor (19, 23), TNF- α -tumor necrosis factor- α (10, 19, 29), 5-HT - 5-hydroxytryptamine (serotonin) (30), ATP - adenosine 5'-triphosphate (24, 25), A - adenosine (24, 25).

(35). These may have some therapeutic potentials for diseases of the male reproductive organs (1, 30, 36, 37).

ACKNOWLEDGEMENTS

- We thank Ruzha Pancheva, B Sc and Nina K. Kresteva, BSc for the valuable discussions.

REFERENCES

- Davidoff MS, Middendorff R, Holstein AF. Dual nature of Leydig cells of the human testis. *BiomedRev* 1996; 6: 11-41
- Jegou B, Sharpe RM. Paracrine mechanisms in testicular control. In: *Molecular Biology of the Male Reproductive System*. Academic Press, New York, London, 1993; 271-310
- Reichlin S. Neuroendocrine-immune interactions. *New EnglJMed* 1993; 329: 1246-1253
- Rivest S, Rivier C. The role of corticotropin-releasing factor and interleukin-1 in the regulation of the neurons controlling reproductive functions. *EndocrRev* 1995; 16: 177-199
- Lyons PD, Blalock JE. The kinetics of ACTH expression in rat leukocyte subpopulations. *JNeuroimmunol* 1995; 63: 103-112
- Besedovsky HO, del Rey A. Immune-neuro-endocrine interactions: facts and hypotheses. *Endocr Rev* 1996; 17: 64-102
- Olsen NJ, Kovacs WJ. Gonadal steroids and immunity. *EndocrRev* 1996; 17: 369-384
- Blalock JE. The syntax of immune-neuroendocrine communications. *Immunol Today* 1994; 15: 504-511
- Hales DB. Leydig cell-macrophage interactions: an overview. In: Payen AH, Hardy MP, Russell LD, editors. *The Leydig Cell*. Cache River Press, Vienna EL, 1996; 453-465
- Xiong Y, Hales DB. Immune-endocrine interaction in the mouse testis: cytokine-mediated inhibition of Leydig cell steroidogenesis. *Endocr inolJ* 1994; 2: 223-228
- Gaytan F, Bellido C, Morales C, Reymundo C, Aguilar E, van Rooijen N. Selective depletion of testicular macrophages and prevention of Leydig cell population after treatment with ethylene dimethane sulfonate in rats. *J ReprodFertil* 1994; 101: 175-182
- Gaytan F, Bellido C, Aguilar E, van Rooijen N. Pituitary-testicular axis in rats lacking testicular macrophages. *Eur JEndocrinol* 1995; 132: 218-222
- Gaytan F, Bellido C, Aceitero J, Aguilar E, Sanchez-Criado J. Leydig cell involvement in the paracrine regulation of mast cells in the testicular interstitium of the rat. *BiolReprod* 1990; 43: 665-671
- Gaytan F, Bellido C, Carrera G, Aguilar E. Differentiation of mast cells during postnatal development of neonatally estrogen-treated rats. *Cell Tissue Res* 1990; 259: 25-31
- Valchanov K, Tonchev A, Pancheva R, Andonov M, Chaldakov GN. Cardiac mast cell proliferation: response to estradiol treatment of neonatal rats [abstract]. *Eur J Morphol* 1997; In press
- Tuncel S, Giirer F, Aral E, Uzuner K, Aydin Y, Baycu C. The effect of vasoactive intestinal peptide (VIP) on mast cell invasion/degranulation in testicular interstitium of immobilized+cold stressed and p-endorphin-treated rats. *Peptides* 1996; 17: 817-824
- Tungel N. Mast cells, vasoactive intestinal peptide (VIP), and the hemorrhagic shock: a possible relationship? *BiomedRev* 1993; 2: 37-46
- LeonA, Buriani A, TosoRD, FabbisM, Romanello S, Aloe L. Mast cells synthesize, store, and release nerve growth factor. *Proc NatlAcadSci USA* 1994; 91: 3739-3743
- McKay DM, Bienenstock J. The interaction between mast cells and nerves in the gastrointestinal tract. *Immunol Today* 1994; 15: 533-538
- Niu X-F, Ibbotson G, Kubes P. A balance between nitric oxide and oxidants regulates mast cell-dependent neutrophil-endothelial cell interactions. *CircRes* 1996; 79: 992-999
- Davidoff MS, Middendorff R, Mayer B, Holstein AF. Nitric oxide synthase (NOS-I) in Leydig cells of the human testis. *Arch Histol Cytol* 1995; 58: 17-30
- Kovacs KJ, Foldes A, Vizi ES. *C-kit* ligand (stem cell factor) affects neuronal activity, stimulates pituitary-adrenal axis and prolactin secretion in rats. *J Neuroimmunol* 1996; 65: 133-141
- Rudge JS, Eaton MJ, Mather P, Lindsay RM, Whittemore SR. CNTF induces raphe neuronal precursors to switch from a serotonergic to a cholinergic phenotype *in vitro*. *Mol CellNeurosci* 1996; 7: 204-221

24. Chaldakov GN, Ghenev PI, Andonov M, Valchanov K, Tonchev A, Pancheva R. Neural-immune-effector (NIE) cross-talk in vascular trophobiology: proposal for new and not yet exploited purinergic regulatory mechanisms. *BiomedRev* 1994; 3: 81-86
25. Neary JT, Rathbone MP, Cattabeni F, Abbracchio MP, Burnstock G. Trophic actions of extracellular nucleotides and nucleosides on glial and neuronal cells. *Trends Neurosci* 1996; 19: 13-18
26. Subramanian N, Bray MA. Interleukin-1 releases histamine from human basophils and mast cells *in vitro*. *J Immunol* 1987; 138: 271-275
27. Konakchieva R, Kyurkchiev S, Kehayov I, Taushanova P, Kanchev L. Selective effect of methoxyindoles on the lymphocyte proliferation and melatonin binding to activated human lymphoid cells. *J Neuroimmunol* 1995; 63: 125-132
28. Elenkov IJ, Hasko G, Kovacs KJ, Vizi ES. Modulation of lipopolysaccharide-induced tumor necrosis factor- α production by selective α - and β -adrenergic drugs in mice. *J Neuroimmunol* 1995; 61: 123-131
29. Soliven B, Albert J. Tumor necrosis factor modulates the inactivation of catecholamine secretion in cultured sympathetic neurons. *J Neurochem* 1992; 58: 1073-1078
30. Dufau ML, Tinajero JC, Fabbri A. Corticotropin-releasing factor: an antireproductive hormone of the testis. *FASEB J* 1993; 7: 229-307
31. Stridsberg M, Fabiani R, Lukinius A, Ronquist G. Prostatomes are neuroendocrine-like vesicles in human semen. *Prostate* 1996; 29: 287-295
32. Harper GP, Barde YA, Burnstock G, Carstairs JR, Dennison ME, Suda K *et al*. Guinea pig prostate is a rich source of nerve growth factor. *Nature* 1979; 279:160-162
33. Hofmann H-D, Unsicker K. The seminal vesicle of the bull: a new and very rich source of nerve growth factor. *Eur J Biochem* 1982; 128:421-426
34. Schultz REH. Regulatory molecules in rat testis. *Acta Universitatis Tamperensis*, Tampere, Finland, 1995; ser A, 434: 5-58
35. Tomov D. Analysis of scientific communications in the field of Sertoli and Leydig cells of the testis [abstract]. Jubilee Scientific Session of the Medical University of Pleven, Pleven 1994; 35 (in Bulgarian)
36. Schill WB, Schneider J, Ring J. The use of ketotifen, a mast cell blocker, for treatment of oligo- and asthenozoospermia. *Andrologia* 1986; 18: 570-573
37. Nagai T, Takaba H, Miyake K, Hirabayashi Y, Yamada K. Testicular mast cell heterogeneity in idiopathic male infertility. *FertilSteril* 1992; 57: 1331-1336

For correspondence:

Dr Kamen P. Valchanov
 Department of Oral Pathology
 King's College
 The Rayne Institute
 123 Coldharbour Lane
 London SE59NU
 United Kingdom

Tel: 44 (171) 3463019

Fax: 44 (171) 3463019

E-mail: RCHVC02@BAY.CC.KCL.AC.UK