

EDITORIAL

ROLE OF ADVENTITIA IN VASCULAR REMODELING IN HYPERTENSION: A TROPHOBIOLOGICAL VIEW

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• The vascular wall has the capacity to undergo remodeling in response to long-term changes or injuries. This is a process of structural rearrangement that involves cell growth, cell death, cell migration, cell modulation and secretion/degradation of extracellular matrix molecules (1). Vascular remodeling is an adaptive phenomenon, e.g. Glagov's compensatory enlargement in atherosclerosis (2), but it may grow into vascular diseases (1), such as hypertension (3), atherosclerosis (4,5), and coronary restenosis after angioplasty (2,6, 7). Nowadays paradigms defining the cell biology of vascular diseases are the following: (*I*) the hypertensive vessel is characterized by hyperinnervation-associated *medial thickening* due to smooth muscle cell (SMC) hypertrophy/hyperplasia and increased extracellular matrix content, (*II*) the atherosclerotic plaque is characterized by SMC/immune cells/increased extracellular matrix-containing *intimal thickening*, and (*III*) the restenotic coronary artery is characterized by SMC/immune cells-containing *neointimal thickening*. The spontaneously hypertensive rats (SHR), the stroke-prone SHR (SHRSP), the genetically hypertensive (GH) rats, and other genetically hypertensive strains are widely used as a model of human essential hypertension. In this volume of *Biomedical Reviews*, Bell (8) updates the knowledge about vascular wall neurotrophobiology in relation to the pathogenesis of hypertension in SHR and GH rats. Also, Kondo *et al* (9) systematize the perivascular nerve-related SMC structural changes in the development of hypertension in SHR and

SHRSP. The data presented in these reviews are evaluated mainly in terms of Levi-Montalcini's neurotrophic theory (10).

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In this context, the role of immune cells in neural-effector interactions should also be considered (see 8 and Refs 23,113-120 therein). The neurobiology of immune cells is one of the expanding fields of neuroscience research, largely due to the involvement of cytokines and neurotrophins in neuroimmune interactions (11). Besides the neurotrophic support of specific populations of neurons, there is an increasing evidence that nerve growth factor (NGF) is "not just for neurons" (12), e.g. it also exerts immunotrophic effects on mast cells and lymphocytes (13, 14). Conversely, stem cell factor (mast cell growth factor, *c-kit* ligand) and leukemia inhibitory factor are not just for immune cells, they also possess neurotrophic effects (11, 15). This *Editorial* will attempt to apply lessons learned from the study of vascular diseases and ageing to the trophobiology of vascular remodeling in hypertension.

• *Lesson from atherosclerosis: do not Ignore adventitial immune cells and perivascular nerves*

In 1962 Schwartz (cited by 16) wrote with respect to the presence of adventitial mononuclear cell infiltration of atheroscle-

rotic vessels: "It is perhaps surprising that such prominent cellular accumulation should have received so little attention... Nevertheless, since cellular infiltration of the adventitia shows such a constant relationship to the presence and degree of plaque formation, it should not be disregarded". This and other related works have been largely ignored (16 and Refs therein), and the atherosclerosis research for a long time has been focused on the intimal changes, appreciating extravasation of immune cells through the arterial lumen, SMC proliferation and hypersecretion of extracellular matrix molecules by SMC (reviewed in 4, 5, 17, in which the word "adventitia" is conceptually absent). However, the observation that adventitial injury alone can lead to intimal thickening is an evidence for the dynamic interaction between the adventitia and intima. Examples of such adventitial injuries include (I) chronic application of lipopolysaccharide (18) and interleukin-1 β (19) to the vascular adventitia, (II) placement of a cuff around the adventitial surface (20), and (Hi) removal of the adventitia (21-23).

Together these studies suggest a potential role of adventitial immune cells and perivascular nerves in atherosclerosis (Fig. 1). Studies aiming at further evaluation of a neural-immune relationship in atherosclerotic adventitia (29-32) are needed.

• *Lesson from coronary restenosis: do not ignore adventitial fibroblasts*

In 1983 at the seminar organized by Dr George Pappas (Department of Anatomy, Medical School, University of Illinois, Chicago, IL), one of us (GNC) delivered a lecture entitled "The fine structure of secretory-state SMC and their possible role in occlusive arterial diseases". During the discussion, the question whether some adventitial fibroblasts may migrate to the intima was raised. The answer of the author was "I do not know. It seems impossible." However, what seemed "impossible" in 1983 was proven possible in 1996 when Shi *et al* (6) and Wilcox and Scott (16) summarized their results indicating that the adventitial fibroblasts proliferate and modulate their phenotype to myofibroblasts migrating to the luminal surface of the balloon-injured coronary arteries, thus contributing to the *neointima formation*. Also, it was recently suggested that *neoadventitial formation*, consisted of fibrotic tissue and mononuclears, could play an important role in coronary restenosis by circumferential neoadventitial, scar-like contraction, which may cause luminal narrowing (7). Collagen type I involvement in cerebral vasospasm (33) may also be at work for adventitial myofibroblast contraction in restenosis. Altogether these data

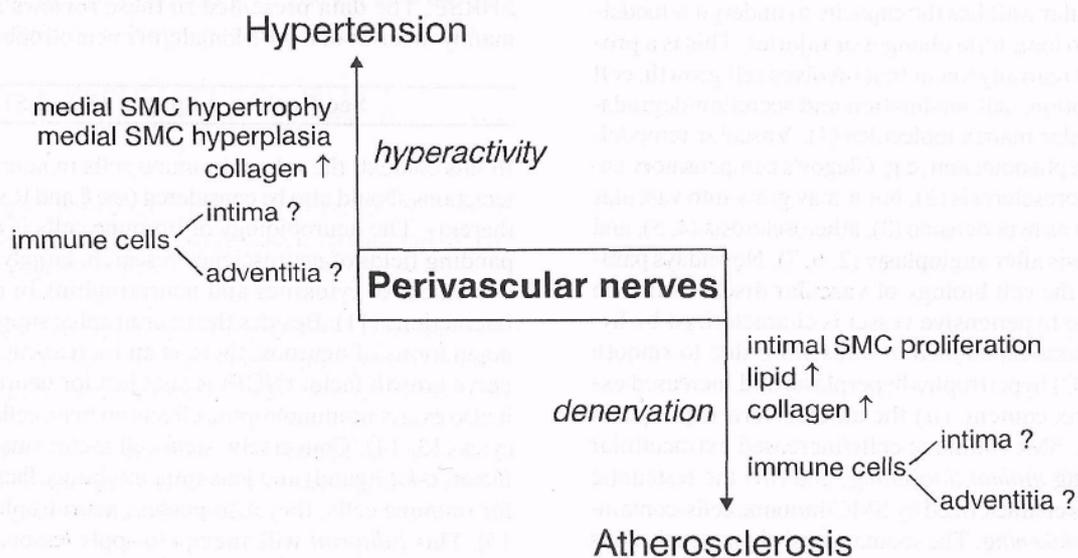


Figure 1. A scheme summarizing data about the role of perivascular nerves in the pathogenesis of hypertension and atherosclerosis, including denervation-associated transplant coronary arteriosclerosis (3, 8, 9, 21-24). Question marks are depicted because of missing data about the role of perivascular nerves for vascular immune cell biology. In other tissues, the denervation results in monocyte invasion into superior cervical ganglion (25), mast cell proliferation in spleen (26), lung (26) and gut (27), and mast cell activation in dura mater (28). SMC - smooth muscle cell.

suggest an important role of adventitial fibroblasts in, and bring into question the sole contribution of SMC to neointimal thickening in coronary restenosis. Moreover, they may bring us back in 1886 to Langhans' description of stellate-shaped cells in atherosclerotic plaques (see 34), thus prompting us to reappraise Virchow's original opinion about the fibroblast nature of these cells. It should also be noted that a large proportion of the intimal foam cells may be derived from foam cells of the media and adventitia rather than from the lumen, after balloon angioplasty of cholesterol-fed rabbits (35).

• ***Lesson from ageing: adventitial immune cells and fibroblasts may also provide a neurotrophic support to perivascular nerves***

Ultrastructurally, perivascular nerve-immune cell (36, 37) and adventitial fibroblast-mast cell (38, 39) links are found in cerebral arteries. Furthermore, there is a correlation between (I) degeneration of SMC, (II) decreased number of adventitial mast cells (see also 40 for human coronary arteries), and (III) atrophy of adventitial fibroblasts (37-39), suggesting that a reduction in the availability of neurotrophins derived from SMC (8, 14), mast cells and/or fibroblasts probably contributes to the decreased density of perivascular nerves during ageing of cerebral arteries (38, 39). These data bring into question the sole contribution of effector SMC to neurotrophic support to perivascular nerves as currently believed.

• ***Implication of the lessons: appreciate the adventitia in hypertension***

In hypertension research, in contrast to atherosclerosis and restenosis, a considerable attention has been paid to the perivascular nerves and their trophic interactions with the medial SMC (3, 8, 9), leaving adventitial immunocytes and fibroblasts ignored. Hence, the morphometry of hypertensive vascular wall traditionally includes measurements of (I) cross-sectional area of intima and media and calculation of media/lumen ratio, the adventitial area being commonly neglected, with an exception of Lee *et al* (42, 43). These authors showed that neonatal sympathectomy alone and its combination with bilateral adrenal demedullation cause adventitial thickening in mesenteric arteries both in SHR and Wistar-Kyoto (WKY) rats. Also, Lee *et al* (43) found a significant increase in the adventitial thickness in SHR compared to that in WKY rats. The importance of the adventitial thickening in SHR arteries and in arteries from sympathectomized animals is unclear. It is known that sympathetic hyperactivity (3, 8, 9) as well as sympathectomy (24) results in collagen overproduction (see Fig. 1). Recent findings show that angiotensin II-induced medial hypertrophy occurs in densely but not sparsely innervated rat arteries (44). These authors suggest a perivascular nerve-mediated hypertrophic effect of angiotensin II. Such a mechanism may also operate in antineurotrophic (9) and/or antiapoptotic (45) action of angiotensin

II in vascular SMC. Note that cardiac mast cell-derived chymase possesses angiotensin I-converting enzyme activity and thus contributes to cardiovascular fibrosis (46). Hence, immune cells in cooperation with medial SMC and with adventitial fibroblasts may be involved in the fibrogenesis of hypertension (see Solomon and Levi-Schaffer [47] in this volume of *Biomedical Reviews* for mast cell-fibroblast interactions in fibrotic diseases, and 48, 49 for immune cells and hypertensive heart fibrosis).

Moreover, evidence is accumulating that bidirectional trophic interactions exist between perivascular nerves and endothelium (50, see also 18-24 mentioned in *Lesson from atherosclerosis*). "It is thus intriguing that loss of substance P (SP) in nerves at the adventitia results in increase in SP at the intima" in the rat pulmonary artery (51). However, results from the same research group "indicate that endothelial substance P is unrelated to the substance P content of sensory nerves since there was no difference in endothelial substance P after capsaicin treatment" in rat mesenteric arteries (52). This could represent a form of intervascular heterogeneity (3). Furthermore, endothelial injury enhances the density of SP-positive perivascular nerves (50). The mechanism by which such an adventitia-endothelium cross-talk operates remains unknown. Possibly, the easily diffusible nitric oxide (53) and carbon monoxide (54), the nexuses between medial SMC, and the adventitial immune cell-derived cytokines carried into the vessel wall by the vasa vasorum could mediate the adventitia-endothelium bidirectional interaction. Until such mediators are not discovered, we suggest they be named *burnstockines*, to appreciate the contribution of Burnstock and coworkers (50, 51).

• ***Conclusion***

The presented data bring into question the sole contribution of vascular SMC to neurotrophic support and, eventually, to medial hypertrophy in hypertensive blood vessels. They raise the significant possibility that a concerted action of different neurotrophins and cytokines derived from multiple cellular sources, i.e. medial SMC, adventitial immune cells and adventitial fibroblasts, is involved in the neurotrophobiology and fibrogenicity of hypertension. This is a substantial part of the hypothesis of neural-immune-effector trophic interactions (38, 39), which application to hypertension research may provide new insights into the pathogenesis and therapy of essential hypertension (Fig.2). In effect, we should recognize that we have paid less attention to the neurotrophic and fibrogenic potentials of the adventitia in vascular remodeling in hypertension. Perhaps it is time to change that.

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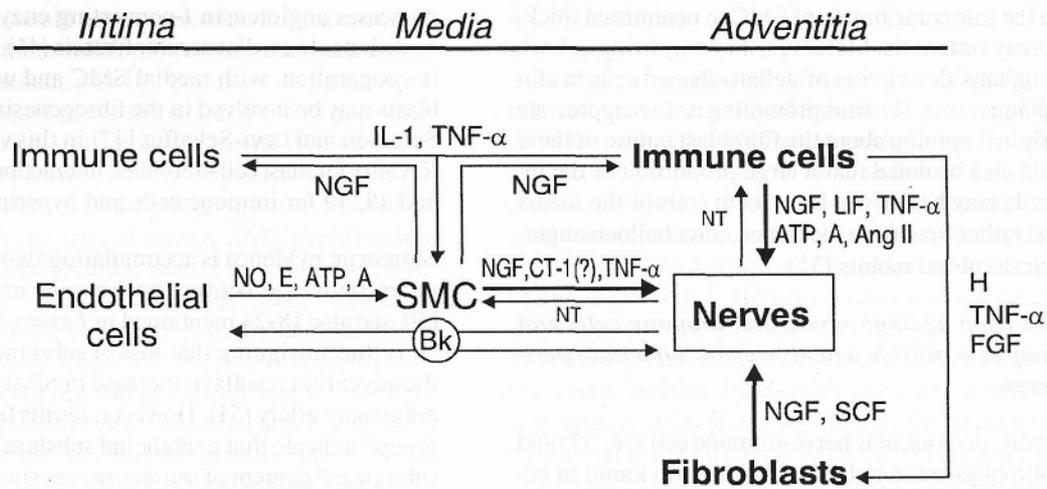


Figure 2. Suggested role of adventitia-based trophic interactions in vascular remodeling in hypertension. Note that three cell types may provide multifactorial neurotrophic support (arrows in bold).

Abbreviations and the respective references: *IL-1* - interleukin-1 (19, 55), *TNF- α* -tumor necrosis factor- α (11, 47, 55), *NGF*-nerve growth factor (11-14), *NO* - nitric oxide (50), *E* - endothelins (50), *ATP* - adenosine S'-triphosphate (56), *A* - adenosine (56), *CT-1* - cardiotrophin-1 (57), *AngII* - angiotensin II (3, 44, 46), *NT* - neurotransmitters (3, 8, 9, 11), *Bk* - burstostockines (50, 51), *LIF* - leukemia inhibitory factor (11), *SCF* - stem cell factor (mast cell growth factor) (15), *H* - histamine (47), *FGF* - fibroblast growth factor (47). Additional mediator molecules may also be involved but are not shown.

REFERENCES

- Gibbons GH, Dzau VJ. The emerging concept of vascular remodeling. *NewEnglJMed* 1994; 330: 1431-1438
- Glagov S. Intimal hyperplasia, vascular modeling, and the restenosis problem. *Circulation* 1994; 89: 2888-2891
- Daemen MJAP, De Mey JGR. Regional heterogeneity of arterial structural changes. *Hypertension* 1995;25(part 1): 464-473
- Yokota T, Hansson GK. Immunological mechanisms in atherosclerosis. *JIntern Med* 1995; 238: 479-489
- Ross R. Cell biology of atherosclerosis. *Annu Rev Physiol* 1995; 57: 791-804
- Shi Y, O'Brien JE, Fard A, Mannion JD, Wang D, Zalewski A. Adventitial myofibroblasts contribute to neointimal formation in injured porcine coronary arteries. *Circulation* 1996; 94: 1655-1664
- Andersen HR, Maeng M, Thorwest M, Falk E. Remodeling rather than neointimal formation explains luminal narrowing after deep vessel wall injury. Insights from a porcine coronary (re)stenosis model. *Circulation* 1996; 93: 1716-1724
- Bell C. Neurotrophic abnormalities and development of high blood pressure in genetically hypertensive rats. *BiomedRev* 1996; 6: 43-55
- Kondo M, Tenkova T, Fujiwara T. Dual effect of sympathetic hyperfunction on blood vessels in spontaneously hypertensive and stroke-prone spontaneously hypertensive rats. *BiomedRev* 1996; 6: 57-68
- Levi-Montalcini R. The nerve growth factor 35 years later. *Science* 1987; 237: 1154-1162
- Rothwell NJ, Hopkins SJ. Cytokines and the nervous system II: actions and mechanisms of action. *Trends Neurosci* 1995; 18: 130-136
- Scully JL, Otten U. NGF: not just for neurons. *Cell Biol Int* 1995; 19: 459-469
- Aloe L, Levi-Montalcini R. Mast cells increase in tissues of neonatal rats injected with the nerve growth factor. *Brain Res* 1977; 133: 358-366
- Braccilaudiero L, Aloe L, Stenfors C, Tirassa P, Theo-

- dorsson E, Lundberg T. Nerve growth factor stimulates production of neuropeptide Y in human lymphocytes. *Neuroreport* 1996; 7: 485-488
15. Carnahan JF, Patel DR, Miller JA. Stem cell factor is a neurotrophic factor for neural crest-derived chick sensory neurons. *JNeurosci* 1994; 14: 1433-1440
 16. Wilcox JN, Scott NA. Potential role of the adventitia in arteritis and atherosclerosis. *Int J Cardiol* 1996; 54 (Suppl): S21-S35
 17. Chaldakov GN, Vankov VN. Morphological aspects of secretion in the arterial smooth muscle cell, with special reference to the Golgi complex and microtubular cytoskeleton. *Atherosclerosis* 1986; 61: 175-192
 18. Prescott MF, McBride CK, Court M. Development of intimal lesions after leukocyte migration into the vascular wall. *AmJPathol* 1989; 135: 835-846
 19. Shimokawa H, Ito A, Fukumoto Y, Kadokami T, Nakaike R, Sakata M *et al*. Chronic treatment with interleukin-1 (3 induces coronary intimal lesions and vasospastic responses in pigs *in vivo*: the role of platelet-derived growth factor. *JClinInvest* 1996; 97: 769-776
 20. Hagihara H, Nomoto A, Mutoh S, Yamaguchi I, Ono T. Role of inflammatory responses in initiation of atherosclerosis: effects of anti-inflammatory drugs on cuff-induced leukocyte accumulation and intimal thickening of rabbit carotid artery. *Atherosclerosis* 1991; 91: 107-116
 21. Booth RFC, Martin JF, Honey AC, Hassall DG, Beesley JE, Moncada S. Rapid development of atherosclerotic lesions in the rabbit carotid artery induced by perivascular manipulation. *Atherosclerosis* 1989; 76: 257-268
 22. Scott TM, Honey AC, Martin JF, Booth RF. Perivascular innervation is lost in experimental atherosclerosis. *Cardioscience* 1992; 3: 145-153
 23. Barker SG, Tilling LC, Miller GC, Beesley JE, Fleetwood G, Stavri GT *et al*. The adventitia and atherogenesis: removal initiates intimal proliferation in the rabbit which regresses on generation of a "neoadventitia". *Atherosclerosis* 1994; 105: 131-144
 24. Fronck K. Trophic effect of the sympathetic nervous system on vascular smooth muscle. *Ann Biomed Eng* 1983; 11: 607-615
 25. Schreiber RC, Shadiack AM, Bennett TA, Sedwick CE, Zigmond RE. Changes in the macrophage population of the rat superior cervical ganglion after postganglionic nerve injury. *JNeurobiol* 1995; 27: 141-153
 26. Dimitriadou V, Rouleau A, Tuong MDT, Newlands GJF, Miller HRP, Luffau G *et al*. Functional relationship between mast cells and C-sensitive nerve fibres evidenced by histamine H₁-receptor modulation in rat lung and spleen. *ClinSci* 1994; 87: 151-163
 27. Osinski MA, Dahl JL, Bass P. Proliferation of mast cells in the smooth muscle of denervated rat jejunum. *JAutonom NervSyst* 1993; 45: 164-174
 28. Bergerot A, Delepine L, Aubineau P. Sympathectomy induces C-fiber sprouting and mast cell activation in the rat dura mater [abstract]. *Eur J Neurosci* 1996; (Suppl 9): 96
 29. Kohchi K, Takebayashi S, Hiroki T, Nobuyoshi M. Significance of adventitial inflammation of the coronary artery in patients with unstable angina: result at autopsy. *Circulation* 1985; 71: 709-716
 30. Baroldi G, Silver MD, Mariani F, Giuliano G. Correlation of morphological variables in the coronary atherosclerotic plaque with clinical patterns of ischemic heart disease. *Am J Cardiovasc Pathol* 1988; 2: 159-172
 31. Chaldakov GN, Valchanov K, Tonchev A, Ghenev PI. The association of mast cells and atherosclerosis: new insights into mast cells in atherogenesis [letter]. *Human Pathol* 1995; 26: 1286
 32. Ghenev P, Valchanov K, Tonchev A, Pancheva R, Chaldakov G. Neural-immune links: an emerging concept of adventitial role in atherogenesis. *Proc Int Med Ass Bulg* 1996; 2: In press
 33. Onoda K, Ono S, Ogihara K, Shiota T, Asari S, Ohmoto T *et al*. Role of extracellular matrix in experimental vasospasm. Inhibitory effect of antisense oligonucleotide on collagen induction. *Stroke* 1996; 27: 2102-2109
 34. Chaldakov GN, Nabika T, Nara Y, Yamori Y. Cyclic AMP- and cytochalasin B-induced arborization in cultured aortic smooth muscle cells: its cytopharmacological characterization. *Cell Tissue Res* 1989; 255: 435-442
 35. Gertz SD, Gimple LW, Ragosta M, Roberts WC, Haber HL, Powers ER *et al*. Response of femoral arteries of cholesterol-fed rabbits to balloon angioplasty with or without laser: emphasis on the distribution of foam cells. *ExpMol Pathol* 1993; 59: 225-243
 36. Dimitriadou V, Aubineau P, Taxi J, Seylaz J. Ultrastructural evidence for a functional unit between nerve fibers and type II cerebral mast cells in the cerebral vascular wall. *Neuroscience* 1987; 22: 621-630
 37. Chaldakov GN, Andrews T, Burnstock G, Cowen T. Neu-

- ral-immune-effector trophobiological links at adventitia-mediaborder in cerebral vessels [abstract]. *Atherosclerosis* 1995; 115 (Suppl): S64
38. Chaldakov GN, Ghenev PI, Andonov M, Valchanov K, Tonchev A, Pancheva R. Neural-immune-effector (NIE) cross-talk in vascular trophobiology: proposal for new and not yet exploited purinergic regulatory mechanisms. *BiomedRev* 1994; 3: 81-86
 39. Chaldakov G. Neural-immune-effector trophic interaction: an emerging concept for the neurotrophic theory [abstract]. *Folia Anat* 1996; 24 (Suppl 2): 24
 40. Kaktursky LV, Yanin VA. Morphometric characteristics of mast cell population in the walls of heart coronary arteries in acute forms of ischemic disease. *Ark Patol* 1994; 6: 71-74
 41. Cowen T. Regulation of the autonomic innervation of blood vessels during development and aging. In: *Vascular Innervation and Receptor Mechanisms: New Perspectives*. Academic Press, New York, London, 1993; 25-40
 42. Lee RMKW, Triggle CR, Cheung DWT, Coughlin MD. Structural and functional consequence of neonatal sympathectomy on the blood vessels of spontaneously hypertensive rats. *Hypertension* 1987; 10: 328-338
 43. Lee RMKW, Borkowski KR, Leenen FHH, Tsoporis J, Coughlin M. Combined effect of neonatal sympathectomy and adrenal demedullation on blood pressure and vascular changes in spontaneously hypertensive rats. *Circ Res* 1991; 69: 714-721
 44. Stassen FRM, Raat NJH, Brouwers-Ceiler DL, Fazzi GE, Smits JFM, De Mey JGR. Angiotensin II induces hypertrophy and hyperreactivity of densely but not sparsely innervated small arteries of the rat. *Vase Res* 1997; In press
 45. Pollman MJ, Yamada T, Horiuchi M, Gibbons GH. Vasoactive substances regulate vascular smooth muscle cell apoptosis. Countervailing influences of nitric oxide and angiotensin II. *CircRes* 1996; 79: 748-756
 46. Ferrario CM, Chappell MC. A new myocardial conversion of angiotensin I. *Curr Opin Cardiol* 1994; 9: 520-526
 47. Solomon A, Levi-Schaffer F. Mast cells beyond allergy: their role in fibrotic conditions. *BiomedRev* 1996; 6: 69-74
 48. Panizo A, Mindan FJ, Galindo MF, Cenarruzabeitia E, Hernandez M, Diez J. Are mast cells involved in hypertensive heart disease? *JHypertens* 1995; 13: 1201-1208
 49. Abumiya T, Masuda J, Kawai J, Suzuki T, Ogata J. Heterogeneity in the appearance and distribution of macrophage subsets and their possible involvement in hypertensive vascular lesions in rats. *Lab Invest* 1996; 75: 125-136
 50. Lincoln J, Ralevic V, Burnstock G. Neurohumoral substances and the endothelium. In: Rubanyi GM, editor. *Cardiovascular Significance of Endothelium-Derived Vasoactive Factors*. Futura Publ, Mount Kisco, New York, 1991; 83-110
 51. Milner P, Loesch A, Burnstock G. Neonatal sensory denervation affects the expression of endothelial peptides in the adult rat pulmonary artery: more cells contain substance P and less contain endothelin. *Endothelium* 1996; 4:71-76
 52. Ralevic V, Dikranian K, Burnstock G. Long-term sensory denervation does not modify endothelial function or endothelial substance P and nitric oxide synthase in rat mesenteric arteries. *J Vase Res* 1995; 32: 320-327
 53. Xiao I, Pang PK. Activation of nitric oxide synthesis in vascular smooth muscle cells and macrophages during development in spontaneously hypertensive rats. *Am J Hypertens* 1996; 9: 377-384
 54. Johnson RA, Colombari E, Lavesa M, Colombari DSA, Talman WT, Nasjletti A. Endogenous carbon monoxide acts on the NTS to lower blood pressure [abstract]. *Hypertension* 1996; 28: 525
 55. De Kimpe S. Leukocytes, cytokines and hypertension; an *in vitro* approach. PhD Thesis, Utrecht University, Utrecht, The Netherlands, 1993
 56. Neary JT, Rathbone MP, Cattabeni F, Abbracchio MP, Burnstock G. Trophic actions of extracellular nucleotides and nucleosides on glial and neuronal cells. *Trends Neurosci* 1996; 19: 13-18
 57. Pennica D, Wood WI, Chien KR. Cardiotrophin-1: a multifunctional cytokine that signals via LIF receptor-gp130 dependent pathways. *Cytokine Growth Factor Rev* 1996; 7: 81-91

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